

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION  
International Bureau



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification <sup>6</sup> :  C12N 9/00		A2	(11) International Publication Number: <b>WO 98/37181</b>  (43) International Publication Date: 27 August 1998 (27.08.98)															
 (21) International Application Number: PCT/US98/03404  (22) International Filing Date: 20 February 1998 (20.02.98)		 (81) Designated States: CA, US, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).  <b>Published</b> <i>Without international search report and to be republished upon receipt of that report.</i>																
 (30) Priority Data: <table><tr><td>60/038,750</td><td>20 February 1997 (20.02.97)</td><td>US</td></tr><tr><td>60/047,151</td><td>20 May 1997 (20.05.97)</td><td>US</td></tr><tr><td>60/054,549</td><td>1 August 1997 (01.08.97)</td><td>US</td></tr><tr><td>60/055,762</td><td>14 August 1997 (14.08.97)</td><td>US</td></tr><tr><td>60/064,322</td><td>30 October 1997 (30.10.97)</td><td>US</td></tr></table> (71) Applicant (for all designated States except US): WHITEHEAD INSTITUTE FOR BIOMEDICAL RESEARCH [US/US]; Nine Cambridge Center, Cambridge, MA 02142 (US).  (72) Inventors; and (75) Inventors/Applicants (for US only): COUNTER, Christopher, M. [CA/US]; Apartment 5R, 310 Prospect Street, Cambridge, MA 02139 (US). MEYERSON, Mathew [US/US]; 11 Aqueduct Road, Wayland, MA 01778 (US). WEINBERG, Robert, A. [US/US]; 25 Copley Street, Brookline, MA 02146 (US).		60/038,750	20 February 1997 (20.02.97)	US	60/047,151	20 May 1997 (20.05.97)	US	60/054,549	1 August 1997 (01.08.97)	US	60/055,762	14 August 1997 (14.08.97)	US	60/064,322	30 October 1997 (30.10.97)	US		
60/038,750	20 February 1997 (20.02.97)	US																
60/047,151	20 May 1997 (20.05.97)	US																
60/054,549	1 August 1997 (01.08.97)	US																
60/055,762	14 August 1997 (14.08.97)	US																
60/064,322	30 October 1997 (30.10.97)	US																
 (74) Agents: GRANAHAN, Patricia et al.; Hamilton, Brook, Smith & Reynolds, P.C., Two Militia Drive, Lexington, MA 02173 (US).																		
 (54) Title: TELOMERASE CATALYTIC SUBUNIT GENE AND ENCODED PROTEIN  (57) Abstract  Isolated DNA encoding the catalytic subunit of a eukaryotic telomerase holoenzyme, such as the catalytic subunit of a yeast or human telomerase holoenzyme; the RNA transcript, which is expressed in primary human tumors, cancer cell lines and telomerase-positive tissues; and the encoded catalytic subunit protein. Methods of assessing cells for malignancy or an increased likelihood of progression to malignancy and methods of diagnosing or aiding in the diagnosis of development of malignancy in an individual are also described.																		

***FOR THE PURPOSES OF INFORMATION ONLY***

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece	ML	Mali	TR	Turkey
BG	Bulgaria	HU	Hungary	MN	Mongolia	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MR	Mauritania	UA	Ukraine
BR	Brazil	IL	Israel	MW	Malawi	UG	Uganda
BY	Belarus	IS	Iceland	MX	Mexico	US	United States of America
CA	Canada	IT	Italy	NE	Niger	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NL	Netherlands	VN	Viet Nam
CG	Congo	KE	Kenya	NO	Norway	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NZ	New Zealand	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	PL	Poland		
CM	Cameroon	KR	Republic of Korea	PT	Portugal		
CN	China	KZ	Kazakhstan	RO	Romania		
CU	Cuba	LC	Saint Lucia	RU	Russian Federation		
CZ	Czech Republic	LI	Liechtenstein	SD	Sudan		
DE	Germany	LK	Sri Lanka	SE	Sweden		
DK	Denmark	LR	Liberia	SG	Singapore		
EE	Estonia						

-1-

TELOMERASE CATALYTIC SUBUNIT GENE AND  
ENCODED PROTEIN

FUNDING

Work described herein was funded, in whole or in part,  
5 by Grant No. R35CA39826 and Grant No. OIG CA 39826 from the  
National Cancer Institute. The U.S. Government has certain  
rights in the invention.

RELATED APPLICATIONS

This application claims the benefit of U.S.  
10 Provisional Application No. 60/038,750, filed February 20,  
1997, entitled "Identification of a Protein Subunit of the  
Yeast Telomerase Holoenzyme" by Christopher M. Counter,  
Matthew Meyerson and Robert A. Weinberg; of U.S.  
Provisional Application No. 60/047,151, filed May 20, 1997,  
15 entitled "Human Telomerase Catalytic Subunit" by  
Christopher M. Counter and Matthew Meyerson; of U.S.  
Provisional Application No. 60/054,549, filed August 1,  
1997, entitled "hEST2, the Putative Human Telomerase  
Catalytic Subunit Gene Is Up-Regulated in Tumor Cells and  
20 During Immortalization" by Matthew Meyerson, Christopher M.  
Counter and Robert A. Weinberg; of U.S. Provisional

- 2 -

Application No. 60/055,762, filed August 14, 1997, entitled "Human Telomerase Catalytic Subunit Gene and Uses Therefor" by Matthew Meyerson, Christopher M. Counter and Robert A. Weinberg; and of U.S. Provisional Application No. 5 60/064,322, filed October 30, 1997, entitled "Human Telomerase Catalytic Subunit Gene and Uses Therefor", by Matthew Meyerson, Christopher M. Counter and Robert A. Weinberg. The entire teachings of these five referenced applications are expressly incorporated herein by 10 reference.

#### BACKGROUND OF THE INVENTION

In most eukaryotes, the telomere ends of linear chromosomes are replicated by the ribonucleoprotein enzyme telomerase (Blackburn, E.H. *Annu. Rev. Biochem.* 53, 163-94 15 (1984); Zakian, V.A., *Science* 270, 1601-7 (1995); Greider, C.W. & Blackburn, E.H., *Cell* 43, 405-13 (1985)). The RNA subunit of this enzyme has now been described in multiple species (Blasco, M.A. et al., *Science* 269, 1267-70 (1995); Feng, J., et al., *Science* 269, 1236-41 (1995); Greider, 20 C.W. & Blackburn, E.H., *Nature* 337, 331-7 (1989); Lingner, J. et al., *Genes Dev.* 8, 1984-98 (1994); McEachern, M.J. & Blackburn, E.H., *Nature* 376, 403-9 (1995); Melek, M. et al., *Mol. Cell Biol.* 14, 7827-38 (1994); Shippen-Lentz, D. & Blackburn, E.H., *Science* 247, 546-52 (1990); Singer, M.S. 25 & Gottschling, D.E., *Science* 266, 404-9 (1994)). Candidate protein subunits have been isolated in the ciliate *Tetrahymena* (Collins, K. et al., *Cell* 81, 677-86 (1995) but not, however, in any genetically tractable organism).

Telomerase is repressed in normal human somatic cells 30 but is re-activated during tumor progression. This re-

-3-

activation is not reflected by changes in the levels of previously cloned genes encoding telomerase subunits.

#### SUMMARY OF THE INVENTION

Described herein are genes which are required for 5 telomerase enzymatic activity and the respective encoded messenger RNAs (transcripts) and proteins. As described below, the present invention relates to DNA which is involved in telomere length regulation in eukaryotes and proteins which are physically associated with the 10 respective active telomerase enzyme and, thus, are each a component of the respective telomerase holoenzyme. In one embodiment, the gene, RNA transcript and encoded protein are a yeast gene and its encoded RNA transcript and protein. In a second embodiment, the gene, RNA transcript 15 and encoded protein are human. Also described herein is a human cDNA which encodes the human telomerase catalytic protein subunit.

The yeast telomerase gene is present on chromosome XII and its disruption alters telomere maintenance, as 20 demonstrated by the decrease in telomere length in transposon-insertion mutants. The sequence of the gene, referred to herein as *EST2* (SEQ ID NO.: 1), the deduced amino acid sequence of the encoded *Est2* protein, also referred to as *Est2p*, (SEQ ID NO.: 2) and the *hEST2* RNA 25 transcript (SEQ ID NO.: 36) are provided. The DNA has been shown to be essential for telomerase activity in yeast and the encoded protein has been shown, by the methods described herein, to be physically associated with telomerase and a constituent of active telomerase complex 30 in yeast. The yeast telomerase DNA *EST2* is a subject of

-4-

the present invention; this includes isolated DNA comprising DNA selected from the following: DNA of SEQ ID NO.: 1; DNA which is the complement of SEQ ID NO.: 1, DNA which hybridizes to DNA of SEQ ID NO.: 1 or a complement thereof; DNA which localizes to yeast chromosome XII; and DNA which encodes the amino acid sequence of SEQ ID NO.: 2. Isolated yeast Est2p is also the subject of this invention. This includes isolated Est2p of SEQ ID NO.: 2 and Est 2p encoded by EST2 DNA as defined herein.

10       The gene described herein is useful to identify genes encoding telomerase proteins in other eukaryotes, particularly in vertebrates, including mammals and especially humans. All or a portion of the gene described can be used. The encoded portion can be used to produce 15 antibodies (monoclonal or polyclonal) which bind the *Est2* protein and can, in turn, be used to identify corresponding proteins (proteins physically associated with telomerase enzymatic activity) in other eukaryotes.

Also described herein is a human cDNA, which was 20 originally named *hEST2* and has been renamed *hTERT*, that encodes the human telomerase catalytic protein subunit, the encoded *hEST2* RNA and the encoded *hEST2* protein. The human DNA described herein is a human homologue of yeast and ciliate genes which encode telomerase catalytic subunits; 25 it shares significant sequence similarity with the telomerase catalytic subunit genes, yeast *EST2* and *Euplotes p123*. *hEST2* RNA expression reflects telomerase activity. The RNA transcript is expressed in primary human tumors, cancer cell lines and telomerase-positive tissues, but is 30 undetectable in telomerase-negative cell lines and differentiated telomerase-negative tissues. *hEST2* message is absent in pre-crisis, telomerase-negative transformed cells, but is readily detectable in post-crisis,

-5-

telomerase-positive immortalized cells. Taken together, these observations are evidence that the induction of *hEST2* mRNA expression is required for the telomerase activation that occurs during cellular immortalization and tumor progression.

The encoded human protein comprises six of the seven conserved sequence motifs which define the polymerase domains of members of the reverse transcriptase family and also includes the invariant aspartic acid residues required for telomerase enzymatic activity. Although *hEST2* protein comprises such motifs which define polymerase domains, beyond these it shows no sequence similarity with reverse transcriptases. It is more closely related to the telomerase catalytic subunits of yeast and ciliates than to other reverse transcriptases. In its domains that lie N-terminal to the polymerase domain, *hEST2* shows clear relatedness to both p123 and Est2p. Many of the sequence identities in the N-termini of the three proteins are in a region just before motif 1. These sequences do not appear in reverse transcriptases or in other proteins and, thus, appear to be unique to telomerases. For example, these three include a unique motif, referred to as the telomerase motif; in *hEST2* this motif extends from amino acid residue 556 to amino acid residue 565 of SEQ ID NO.: 3, with absolute invariant sequence extending from amino acid residue 560 to amino acid residue 565 of SEQ ID NO.: 3. Further, within the *hEST2* domain that shares sequence similarity with reverse transcriptases, it is clear that *hEST2* is more closely related to telomerase reverse transcriptases than to non-telomerase reverse transcriptases. The three telomerase catalytic subunits (*hEST2*, yeast EST2 and *Euplotes* p123) form a subgroup within the reverse transcriptase family.

-6-

The present invention relates to isolated DNA which encodes an RNA transcript which is expressed in primary human tumors, cancer cell line and telomerase-positive tissue. In one embodiment, the isolated DNA comprises 5 *hEST2* DNA such as DNA comprising SEQ ID NO.: 35 (DNA comprising the nucleotide sequence of SEQ ID NO.: 35). Isolated DNA of the present invention encodes an RNA transcript which is not detectable in telomerase-negative cell lines or in differentiated telomerase-negative 10 tissues. Further, isolated DNA of the present invention encodes an RNA transcript which is not detectable in pre-crisis, telomerase-negative transformed cells and is detectable in post-crisis, telomerase-positive immortalized 15 cells. In a particular embodiment, the DNA comprises *hEST2* cDNA, such as SEQ ID NO.: 35 or DNA which hybridizes thereto or to a complement of *hEST2* cDNA (such as a complement of SEQ ID NO.: 35) under conditions of high stringency. A further subject of the present invention is 20 an isolated RNA transcript encoded by isolated DNA of the present invention.

A method of altering telomerase function and, thus, of altering telomere shortening is also described. In the method, expression and/or function of the telomerase-associated protein is altered (enhanced or reduced), 25 resulting in altered (enhanced or reduced) cell lifespan. Expression of the telomerase-associated protein is enhanced, for example, by introducing DNA encoding the protein into cells. Alternatively, telomerase-associated protein is introduced into cells. Expression of the 30 telomerase-associated protein is reduced, for example, by introducing into cells an agent which inhibits production or function of the protein, an agent which destroys the expressed protein or a dominant negative form of the

-7-

protein. Therefore, the present invention also provides a method of increasing the lifespan of a cell or, alternatively, of reducing the lifespan of a cell, such as that of cancer cells or transformed cells. In the method 5 of increasing lifespan of cells, telomerase activity is maintained or increased within the cells, such that telomeres are maintained and as the cells age, telomere shortening does not occur or occurs to a lesser extent than would otherwise be the case (if telomerase activity were 10 not maintained or increased). Reduction of lifespan of cells, such as tumor cells, is accomplished by introducing into the cells an inhibitor of the telomerase protein subunit described herein.

A method of assessing cells or aiding in the 15 assessment of cells for malignancy or an increased likelihood of progression to malignancy is also the subject of this invention. In the method, cells to be assessed are obtained from an individual (e.g., a human) in need of such an assessment. The cells are processed in such a manner 20 that DNA, RNA or both in the cells are rendered available for annealing or hybridization with complementary polynucleotides or oligonucleotides (DNA or RNA), such as probes or primers, thereby producing processed cells. The processed cells are combined with DNA or RNA required for 25 telomerase enzymatic activity (DNA or RNA encoding a protein required for telomerase enzymatic activity), a complement of the required DNA or RNA or a characteristic portion or fragment of the DNA or RNA, such as the telomerase motif or all or a portion of the 5' end of *hEST2* 30 which is not shared with the yeast or *Euplotes* gene. If hybridization occurs, it is indicative of the presence of telomerase protein-encoding DNA or RNA in the cells and of activation of telomerase, which is also indicative of malignancy or an increased likelihood of progression to

-8-

malignancy. As described herein, the *hEST2* DNA encodes an RNA transcript which is expressed in primary tumors, cancer cell lines and telomerase - positive tissues and is readily detectable in post-crisis, telomerase-positive immortalized 5 cells, but not detectable in telomerase-negative cell lines and differentiated telomerase-negative tissues. Detection of *hEST2* DNA and/or of the RNA transcript, thus, makes it possible to detect DNA which encodes a transcript required for telomerase activation that occurs during cellular 10 immortalization and tumor progression and to detect the RNA transcript itself.

In an alternative embodiment, the invention is a method of diagnosing or aiding in the diagnosis of development of malignancy (cancer, tumor formation) in an 15 individual, in which the occurrence (presence or absence) and/or quantity of a telomerase protein (e.g., *hEST2* protein) in cells is assessed. In the method, cells are obtained from an individual in need of diagnosis of malignancy and processed, if necessary, in such a manner 20 that proteins in the cell are available for detection, such as by binding with antibodies or antibody fragments. The processed cells are combined or contacted with antibodies that recognize (bind) the telomerase protein (e.g., antibodies that bind Est2p or *hEST2* protein) under 25 conditions appropriate for antibody binding to occur. Whether binding occurs is determined; optionally, the extent to which binding occurs can also be determined. If binding of anti-telomerase protein antibodies (e.g., anti-*hEST2* protein antibodies) to a component of processed cells 30 occurs, it is indicative of the presence of *hEST2* protein in the cells and of malignancy or an increased likelihood of development or malignancy in the individual.

In one embodiment, the present invention is a method of reducing expression of *hEST2* RNA and *hEST2* protein in

- 9 -

cells of an individual (e.g., a human or other mammal). The method comprises administering to the individual a drug selected from the group consisting of drugs which inhibit (directly or indirectly) or bind *hEST2* RNA and prevent or 5 reduce production of *hEST2* protein and drugs which inhibit *hEST2* protein function or activity. The drug is administered in a therapeutically effective amount (an amount sufficient to have the desired effect of reducing expression of *hEST2* RNA and *hEST2* protein) and under 10 conditions appropriate for entry into cells, in which they have the desired effect.

In a further embodiment, the present invention is a method of treating cancer in an individual, in need of such treatment. In the method, a drug which inhibits or binds 15 *hEST2* RNA (or DNA) and prevents or reduces production of *hEST2* protein or a drug which inhibits *hEST2* protein function or activity is administered to the individual under conditions appropriate for entry into cells and in a therapeutically effective amount (an amount sufficient to 20 have the desired effect of inhibiting *hEST2* RNA and/or preventing or reducing production of *hEST2* protein or inhibiting *hEST2* protein function or activity), with the result that cancer in the individual is treated (reduced, reversed or prevented from advancing).

25 A further embodiment of the invention is a method of altering (increasing or reducing) lifespan of cells in culture or in an individual. In the method in which lifespan is increased, *hEST2* DNA is introduced into cells (e.g., in culture or in the individual), in which the *hEST2* 30 RNA transcript and encoded protein are produced in sufficient quantity to increase the lifespan of cells in the cultured cells or the individual. In the method in which lifespan is decreased, *hEST2* protein function,

-10-

activity or production is reduced (partially or totally). This is done, for example, by introducing into cells in culture or administering to an individual in whom cell lifespan is to be decreased a drug which inhibits or binds 5 *hEST2* RNA and prevents or reduces production of *hEST2* protein; a drug which inhibits *hEST2* protein function, activity or production; or a drug which inhibits or binds *hEST2* DNA and prevents or reduces production of *hEST2* RNA transcripts. The drug is administered to an individual 10 under conditions appropriate for entry into cells in sufficient quantity to have the desired effect (in sufficient quantity to decrease lifespan of the cells by reducing *hEST2* protein function, activity or production).

In the above-described embodiments, the drug can be, 15 for example, a small organic molecule, an enzyme which degrades *hEST2* protein, an enzyme inhibitor (e.g., a telomerase or *hEST2* enzyme inhibitor), an *hEST2* transcriptional regulator; an antisense molecule or a dominant negative form of *hEST2* protein.

20 Also described herein is a mutant yeast strain, referred to as DN, which carries an inactive chromosomal *rad52Δ* allele. The RAD 52 coding sequence of a haploid strain of genotype *MATa leu 2-3, 112 lys2-201 trpl-1 ura3-52 his3-200* was replaced with a selectable marker (e.g., 25 the *HIS3* gene), to generate strain DN. Mutant yeast strain DNR, in which the absence of *RAD52* was complemented by introducing the plasmid *pYPCR+<sup>35</sup>*, which encodes the *RAD52* gene under its own promoter and the selectable marker *URA3*, is also described. Also described are mutant yeast strain 30 *DNRtlc4*, in which the *TLC1* gene in DNR cells was replaced with the *LEU2* selectable marker gene and Y0035, in which

-11-

the Est2 gene has been mutated with the addition of a TN3 LEU2 transposon in the DNR genetic background.

The mutant yeast strain described herein requires telomerase activity for viability. It is useful to carry 5 out a rapid and automatable biological assay for telomerase inhibitors and, thus, for inhibitors of telomere biosynthesis. The assay is also the subject of the present invention. The mutant yeast strain is also useful to identify agents which are recombination inhibitors. In the 10 mutant yeast strain, referred to as DN, the *RAD52* recombination gene has been replaced by a *HIS3* gene. As described herein, disruption of the *TLC1* telomerase RNA subunit gene or the *MIT1* putative telomerase protein subunit gene in the *rad52* mutants leads to a delayed cell 15 death. Yeast lacking both recombination activity and telomerase activity die after about 20 to 60 cell generations. Telomerase activity can be disrupted by enzyme inhibitors, as well as by genetic inactivation. Thus, for a telomerase inhibitor screen, test compounds are 20 added, at different concentrations, to the DNA mutant strain (the test strain) and to a wild-type (control) strain. Periodically, such as each day, cultures of the two strains are diluted (e.g., 1:1000). Compounds that kill the yeast instantly or that kill the DNA and wild-type 25 yeast with identical delays, are eliminated. Compounds that kill the test yeast strain significantly before the control yeast strain are telomerase inhibitors. This biological assay has several advantages over available methods. It is fast, automatable, and can be performed 30 easily in large quantities. Because it is a biological assay, it reflects the physiological function of the drugs. Furthermore, it allows testing of biochemical and uptake parameters in a single step. Finally, this assay also

-12-

excludes inhibitors of other metabolic enzymes, such as RNA and DNA polymerases and is specific for telomerase inhibitors.

#### BRIEF DESCRIPTION OF THE DRAWINGS

5 Figures 1A and 1B present the nucleic acid sequence of the *EST2* gene (SEQ ID NO.: 1) and the predicted amino acid sequence of the Est2 protein (SEQ ID NO. 2).

Figure 2 shows alignment of the predicted amino acid sequence of hEST2, also referred to as *hTERT*, (SEQ ID NO.: 10 3) with the yeast Est2p (SEQ ID NO.: 2) and *Euplotes p123* (SEQ ID NO.: 4) homologues. Amino residues within shaded blocks are identical between at least two proteins. Identical amino acids within the reverse transcriptase (RT) motifs (Poch et al., *EMBO J.* 8:3867-3874 (1989); Xiong and 15 Eickbush, *EMBO J.* 9:3353-3362 (1990)) are in black boxes, an example of a telomerase-specific motif in an outlined gray box, and all other identical amino acids in gray boxes. RT motifs are extended in some cases to include other adjacent invariant or conserved amino acids. The 20 sequence of the expressed sequence tag AA281296 is underlined.

Figures 3A-3F show the alignment of RT motifs 1 to 6 of telomerase subunits hEST2 (SEQ ID NO.: 5-10), p123 (SEQ ID NO.: 11-16) and Est2p (SEQ ID NO.: 17-22) with *S. cerevisiae* group II intron-encoded RTs a2-Sc (SEQ ID NO.: 23-28) and a1-Sc (SEQ ID NO.: 29-34). The consensus sequence of each RT motif is shown (h: hydrophobic residues, p: small polar residues, c: charged residues). Amino acids that are invariant among telomerases and the RT 30 consensus are boxed in black, while those that are invariant among telomerases and similar to the RT consensus are boxed in gray. Open boxes identify highly conserved

-13-

residues unique to either telomerases or to non-telomerase RTs. Asterisks denote amino acids essential for polymerase catalytic function.

Figure 4 is an ideogram of human chromosome 5p showing linkage of *hEST2* to sequence-tagged site (STS) markers WI-9907 and D5S417 determined by radiation hybrid (RH) mapping employing the Genebridge 4 RH panel (GB) (1 cR=270 kb). The higher resolution Stanford G3 RH panel (G3) placed *hEST2* close to marker D5S678 (1 cR=30 kb). The calculated distance between *hEST2* and these STS markers is in centiRays (cR) with LOD scores in parentheses.

Figures 5A-5B show the nucleotide sequence of *hEST2* cDNA in which the start codon ATG is underlined (SEQ ID NO.: 35).

Figure 6 is the amino acid sequence of *hEST2* (SEQ ID NO.: 3).

Figures 7A, 7B and 7C are the nucleotide sequence of partially or alternatively spliced *hEST2* message RNA (SEQ ID NO.: 36).

Figures 8A-8F show the expression of *hEST2* in normal human tissues and cancer cell lines. Duplicate RNA blots were probed with an *hEST2* probe (top panels) or with a  $\beta$ -actin probe as internal control (bottom panels). The 4.4 kb, 6 kb, and 9.5 kb transcripts are *hEST2*-specific.

Figures 9A-9D shows *hEST2* expression in primary human tumors. Rnase protection assays are shown for *hEST2* and  $\beta$ -actin controls. Sizes of the full-length and protected bands are indicated. Shown are the HL60 leukemia cell line (control), normal breast tissue, 2 primary breast tumors, the MCF7 and T47D breast cancer cell lines, and normal and primary tumor tissues from the testis, colon, and ovary. The doublet seen protected by the *hEST2* probe is invariant, and may be a result of probe secondary structure.

-14-

#### DETAILED DESCRIPTION OF THE INVENTION

The linear chromosomes of eukaryotic cells offer the biological advantages of rapid recombination, assortment, and genetic diversification. However, linear DNA is inherently more unstable than circular forms. To address this difficulty, the eukaryotic chromosome has evolved to include a DNA-protein structure, the telomere, which caps chromosome ends and protects them from degradation and end-to-end fusion (Blackburn, *Ann. Rev. Biochem.* 53:163-194 (1984); Zakian, *Science* 270:1601-1607 (1995)).

The DNA component of telomeres consists of tandem repeats of guanine-rich sequences that are essential for telomere function (Blackburn, *Ann. Rev. Biochem.* 53:163-194 (1984), Blackburn, *Nature* 350:569-573 (1991); Zakian, *Science* 270:1601-1607 (1995)). These repeats are replicated by conventional DNA polymerases and by a specialized enzyme, telomerase (Greider, In *Telomeres*, eds. Blackburn and Greider, Cold Spring Harbor Laboratory Press, pp. 35-68 (1995)), first identified in the ciliate *Tetrahymena* (Greider and Blackburn, *Cell* 43:405-413 (1985)). The telomerase enzyme is essential for complete replication of telomeric DNA because the cellular DNA-dependent DNA polymerases are unable to replicate the ultimate ends of the telomeres due to their requirement for a 5' RNA primer and their unidirectional mode of synthesis. Removal of the most terminal RNA primer following priming of DNA synthesis leaves a gap that cannot be replicated by these polymerases (Olovnikov, *Dokl. Akad. Nauk SSSR* 201:1496-1499 (1971); Watson, *Nature New Biol.* 239:197-201 (1972)). Telomerase surmounts this problem by *de novo* addition of single-stranded telomeric DNA to the ends of chromosomes (Greider and Blackburn, *Cell* 43:405-413 (1985),

-15-

Greider and Blackburn, *Nature* 337:331-337 (1989); Yu et al., *Nature* 344:126-132 (1990); Greider, In *Telomeres*, eds. Blackburn and Greider, Cold Spring Harbor Laboratory Press, pp. 35-68 (1995)).

- 5       The telomerase enzymes that have been characterized to date are RNA-dependent DNA polymerases which synthesize the telomeric DNA repeats using an RNA template that exists as a subunit of the telomerase holoenzyme (Greider, In *Telomeres*, eds. Blackburn and Greider, Cold Spring Harbor
- 10      Laboratory Press, pp. 35-68 (1995)). The genes specifying the RNA subunits of telomerases have been cloned from a wide variety of species, including humans (Feng et al., *Science* 269:1236-1241 (1995); Greider, In *Telomeres*, eds. Blackburn and Greider, Cold Spring Harbor Laboratory Press,
- 15      pp. 35-68 (1995)) and been shown in several instances to be essential for telomerase function *in vivo* (Yu et al., *Nature* 344:126-132 (1990); Yu and Blackburn, *Cell* 67:823-832 (1991); Singer and Gottschling, *Science* 266:404-409 (1994); Cohn and Blackburn, *Science* 269:396-400 (1995);
- 20      McEachern and Blackburn, *Nature* 376:403-409 (1995)). In addition, three proteins have been identified to date that are associated with telomerase activity. p80 and p95 were purified from the ciliate *Tetrahymena* (Collins et al., *Cell* 81:677-686 (1995)) and the gene encoding a mammalian homologue of p80, TP1/TLP1, has also been cloned (Harrington et al., *Science* 275:973-977 (1997); Nakayama et al., *Cell* 88:875-884 (1997)). The specific mechanism by which these proteins participate in telomerase function has not been defined.
- 25      Perturbing the function of telomerase in the ciliate *Tetrahymena*, through the overexpression of an inactive form

-16-

of the telomerase RNA, or in yeast, through the mutation of genes encoding either the catalytic protein or template RNA subunit, leads to progressive telomere shortening as cells pass through successive cycles of replication (Yu et al., 5 *Nature* 344:126-132 (1990); Singer and Gottschling, *Science* 266:404-409 (1994); McEachern and Blackburn, *Nature* 376:403-409 (1995); Lendvay et al., *Genetics* 144:1399-1412 (1996); Counter et al., *Proc. Natl. Acad. Sci., USA* 94:9202-9207 (1997); Lingner et al., *Science* 276:561-567 10 (1997)). This loss of telomeric DNA is ultimately lethal if it is not overcome. The lethality seems to be triggered when telomeres have been truncated below a critical threshold level. Hence, in the absence of compensating mechanisms, yeast cell lineages that lack telomerase 15 activity have a lifespan dictated by the lengths of their telomeres.

In humans, telomerase activity is readily detectable in germline cells and in certain stem cell compartments. However, enzyme activity is not detectable in most somatic 20 cell lineages (Harley et al., *Cold Spring Harb. Symp. Quant. Biol.* 59:307-315 (1994); Kim et al., *Science* 266:2011-2015 (1994); Broccoli et al., *Proc. Natl. Acad. Sci., USA* 92:9082-9086 (1995); Counter et al., *Blood* 85:2315-2320 (1995); Hiyama et al., *J. Immunol.* 155:3711-25 3715 (1995)). Consistent with this, telomeres of most types of human somatic cells shorten with increasing organismic age and with repeated passaging in culture, similar to the situation seen in protozoan and yeast cells that have been deprived experimentally of a functional 30 telomerase enzyme (Harley et al., *Nature* 345:454-460 (1990); Hastie et al., *Nature* 346:866-868 (1990)).

-17-

Eventually the proliferation of cultured human cells will halt at a point termed senescence (Hayflick and Moorhead, *Exp. Cell. Res.* 25:585-621 (1961); Goldstein, *Science* 249:1129-1133 (1990)), apparently before the telomeres of 5 these cells have become critically short.

Cultured normal human cells can circumvent senescence and thereby continue to proliferate when transformed by a variety of agents. In such cultures, telomere shortening continues until a subsequent point is reached that is 10 termed crisis, where telomeres have become extremely short (Counter et al., *EMBO J.* 11:1921-1929 (1992) and *J. Virol.* 68:3410-3414 (1994); Shay et al., *Oncogene* 8:1407-1413 (1993); Klingelhutz et al., *Mol. Cell. Biol.* 14:961-969 (1994). Crisis, perhaps best described in SV-40 15 transformed cells, is characterized by karyotypic instability, particularly the types of instability observed in chromosomes lacking functional telomeres, and by significant levels of cell death (Sack, *In Vitro* 17:1-19 (1981)). The crisis phenotype is reminiscent of that 20 observed in yeast and Tetrahymena cells in which telomerase function has been experimentally perturbed.

The simplest interpretation of these data is that the lifespan of telomerase-negative human cells, like that of their yeast and ciliate counterparts, is ultimately limited 25 by the length of telomeres. Rare human cells that have acquired the ability to grow indefinitely emerge from crisis populations with a frequency of  $10^{-6}$  to  $10^{-7}$  (Huschtscha and Holliday, *J. Cell Sci.* 63:77-99 (1983); Shay and Wright, *Exp. Cell Res.* 184:109-118 (1989)). This 30 implies that a mutational event is required to confer the immortal phenotype on these cells. The immortal cells that escape crisis are characterized by readily detectable

-18-

levels of telomerase activity and by stable telomeres (Counter et al., *EMBO J.* 11:1921-1929 (1992) and *J. Virol.* 68:3410-3414 (1994); Shay et al., *Mol. Cell. Biol.* 15:425-432 (1995); Whitaker et al., *Oncogene* 11:971-976 (1995); 5 Gollahon and Shay, *Oncogene* 12:715-725 (1996); Klingelhutz et al., *Nature* 380:79-82 (1996)). This suggests that activation of telomerase can overcome the limitations imposed by telomere length on the lifespan of cell lineages.

10 Activation of telomerase also appears to be a major step in the progression of human cancers. Unlike normal human cells, cancer cells can be established as permanent cell lines and, thus, are presumed to have undergone immortalization during the process of tumorigenesis.

15 Moreover, telomerase activity is readily detected in the great majority of human tumor samples analyzed to date (Counter et al., *Proc. Natl. Acad. Sci., USA* 91:2900-2904 (1994); Kim et al., *Science*, 266: 2011-2015 (1994); Shay and Bacchetti, *Eur. J. Cancer* 33:787-791 (1997)).

20 Taken together, these various observations have been incorporated into a model which proposes that the limitation on prolonged cell replication imposed by telomere shortening serves as an important anti-neoplastic mechanism used by the body to block the expansion of 25 pre-cancerous cell clones. According to such a model, tumor cells transcend the crisis barrier and emerge as immortalized cell populations by activating previously unexpressed telomerase, enabling them to restore and maintain the integrity of their telomeres (Counter et al.,

30 *EMBO J.* 11:1921-1929 (1992) and *Proc. Natl. Acad. Sci., USA* 91:2900-2904 (1994); Harley et al., *Cold Spring Harb. Symp. Quant. Biol.* 59:307-315 (1994)).

-19-

A major question provoked by this model is the mechanism used to resurrect telomerase expression during tumor progression. Expression of the telomerase-associated protein TP1/TLP1 does not reflect the level of telomerase activity (Harrington et al., *Science* 275:973-977 (1997); Nakayama et al., *Cell* 88:875-884 (1997)). It is also clear that the levels of the human telomerase RNA component, *hTR*, cannot completely explain the regulation of telomerase activity. Although the levels of *hTR* and its mouse counterpart, *mTR*, increase with tumor progression (Feng et al., *Science* 269:1236-1241 (1995); Blasco et al., *Nat. Genet.* 12:200-204 (1996); Broccoli et al., *Mol. Cell. Biol.* 16:3765-3772 (1996); Soder et al., *Oncogene* 14:1013-1021 (1997)), the amounts of these transcripts do not always correlate with enzymatic activity. Indeed, *hTR* or *mTR* transcript levels can be significantly higher in telomerase-negative cells and tissues than in telomerase-positive cancer cells (Avilion et al., *Cancer Res.* 56:645-650 (1996); Bestilny et al., *Cancer Res.* 56:3796-3802 (1996); Blasco et al., *Nat. Genet.* 12:200-204 (1996)). Similarly, even though telomerase levels increase 100- to 2000-fold during the immortalization of human cells, the level of *hTR* message increases at most two-fold (Avilion et al., *Cancer Res.* 56:645-650 (1996)). Therefore, derepression of the *hTR* and TP1 subunits cannot easily be invoked to explain the appearance of telomerase activity in the great majority of human tumor samples. Thus far, the rate-limiting step in telomerase activation has remained elusive.

Described herein is work which has shown that mutations in a gene of eukaryotic origin (in an open

-20-

reading frame required for telomere maintenance) results in loss (absence) of telomerase enzymatic activity in cells and that the encoded protein is physically associated with the active telomerase enzyme, thus showing that the DNA 5 encodes a component of a eukaryotic telomerase holoenzyme. Also described herein is isolated DNA of eukaryotic origin which is required for telomerase enzymatic activity and the isolated encoded RNA transcripts and proteins. Specifically, *EST2*, a yeast gene, and *hEST2*, which is of 10 human origin and the encoded respective transcripts and proteins (yeast *EST2p* and human *hEST2*) are described, as are their respective sequences. DNA of the present invention includes cDNA, genomic DNA, recombinantly produced DNA and chemically synthesized DNA. It can be 15 obtained from a source in which it occurs in nature or can be produced using known chemical or recombinant DNA methods. The following is a discussion of work carried out relating to yeast and human telomerase protein.

Gene Encoding Protein Subunit Of Yeast Telomerase  
20 Holoenzyme and the Encoded Protein  
As described herein, it has been shown, using a genetic screen in budding yeast, that mutations in *EST2*, an open reading frame required for telomere maintenance, (Lendvay, T.S. et al., *Genetics* 144, 1399-1412 (1996) 25 result in the absence of telomerase enzymatic activity in cells. As also described herein, epitope tagging and immunochemical studies showed that the Est2 protein is physically associated with the active telomerase enzyme. Thus, *EST2* encodes a component of the yeast telomerase 30 holoenzyme.

Interest in telomeres and telomerase has heightened in recent years with the discovery that telomerase is present

-21-

at low, almost undetectable, levels in most human somatic tissues and is readily detectable in germline cells and in the vast majority of tumor cell samples (Counter, C.M. et al., *Proc. Natl. Acad. Sci. USA* 91, 2900-4 (1994); Kim,  
5 N.W. et al., *Science* 266, 2011-5 (1994). Somatic cell lineages which lack telomerase lose telomere segments progressively as they pass through successive replication cycles, limiting their lifespan (Harley, C.B. et al.,  
*Nature* 345, 458-60 (1990), Hastie, N.D. et al., *Nature* 346,  
10 866-8 (1990)). Conversely, cell populations that resurrect telomerase expression ensure maintenance of telomere length, thereby removing a barrier to their further unlimited replication and resulting in the process termed immortalization (Counter, C.M. et al., *Embo. J.* 11, 1921-9  
15 (1992)). These observations have led to a model which proposes that the process of telomere shortening limits the replicative potential of most human somatic cell lineages, and that cancer cells overcome this limitation by activating telomerase expression during the course of tumor  
20 progression (Counter, C.M. et al., *Embo. J.* 11, 1921-9 (1992); Harley, C.B. et al., *Cold Spring Harb. Symp. Quant. Biol.* 59, 307-15 (1994)).

Validation of this model has been held back by the difficulties associated with isolating and characterizing  
25 the telomerase enzyme and its encoding genes. Much of this work has focused on the telomerase enzymes of ciliates; the formation of thousands of mini-chromosomes present in their macro-nuclei seems to require correspondingly high amounts of telomerase, dwarfing the amounts available for study in  
30 metazoan cells. The telomerase enzyme in *Tetrahymena* has been reported to be composed of two proteinaceous and one RNA subunit, the latter responsible for templating the

-22-

telomeric DNA segments (Greider, C.W. & Blackburn, E.H., *Nature* 377, 331-7 (1989); Collins, K. et al., *Cell* 81, 677-86 (1995); Yu, G.L. et al., *Nature* 344, 126-32 (1990)).  
The protein subunits of telomerase have not been described  
5 in any other organism, although the genes specifying the  
RNA subunits of telomerase have been cloned in a wide range  
of other species (Blasco, M.A. et al., *Science* 269, 1967-70  
(1995); Feng, J., et al., *Science* 269, 1236-41 (1995);  
Lingner, J. et al., *Genes Dev.* 8, 1984-98 (1994);  
10 McEachern, M.J. & Blackburn, E.H., *Nature* 376, 403-9  
(1995); Melek, M. et al., *Mol. Cell Biol.* 14, 7827-38  
(1994); Shippen-Lentz, D. & Blackburn, E.H., *Science* 247,  
546-52 (1990); Singer, M.S. & Gottschling, D.E., *Science*  
266, 404-9 (1994)).  
15 Genetic strategies designed to identify and isolate  
telomerase-encoding genes have centered on the yeast, *S.*  
*cerevisiae*. Mutations in several genes have been found to  
lead to progressive telomere shortening. Of these genes,  
*TLC1*, which specifies the RNA subunit of the telomerase  
20 enzyme (Singer, M.S. & Gottschling, D.E., *Science* 266, 404-  
9 (1994)), is required for telomerase enzymatic activity  
(Cohn, M. & Blackburn, E.H., *Science* 269, 396-400 (1995);  
Lin, J.J. & Zakian, V.A., *Cell* 81, 1127-35 (1995); Lue,  
N.F. & Wang, J.C., *J. Biol. Chem.* 270, 21453-6 (1995);  
25 Steiner, B.R. et al., *Proc. Natl. Acad. Sci. USA* 93, 3817-  
21 (1996)). On the other hand, *EST2* encodes a protein of  
unknown function (Lendvay, T.S. et al., *Genetics* 144, 1399-  
1412 (1996)); *EST1* (Lundblad, V. & Szostak, J.W., *Cell* 57,  
633-43 (1989)) and *CDC13* (Garvik, B. et al., *Mol. Cell*  
30 *Biol.* 15, 6128-38 (1995); Nugent, C.I., Hughes, T.R., Lue,

-23-

N.F. & Lundblad, V., *Science* 274, 249-52 (1996)) are not essential for telomerase activity (Cohn, M. & Blackburn, E.H., *Science* 269, 396-400 (1995); Nugent, C.I. et al., *Science* 274, 249-52 (1996)); and *KEM1* appears to encode a 5 nuclease that acts on telomeres (Liu, Z. & Gilbert, W., *Cell* 77, 1083-92 (1994); Liu, Z. et al., *Proc. Natl. Acad. Sci. USA* 92, 6002-6 (1995)). A search for genes encoding telomerase protein subunits was initiated because the protein subunits of the yeast telomerase have not yet been 10 identified, and because there are no obvious homologs of the *Tetrahymena* telomerase protein subunits encoded in the yeast genome (Collins, K. et al., *Cell* 81, 677-86 (1995); Cherry, J.M. et al., *Saccharomyces* Genome Database (<http://genome-www.stanford.edu/Saccharomyces>)).  
15 A genetic screen designed to identify the genes specifying telomerase subunits was undertaken, based on the observation that telomere loss in yeast cells lacking the telomerase RNA component can be compensated by the actions of a second telomere-maintenance system that utilizes the 20 DNA recombination machinery. An essential component of this machinery is the product of the *RAD52* gene. Accordingly, the simultaneous inactivation of *RAD52* and the telomerase RNA component results in cell lethality in both *S. cerevisiae* and *K. lactis* (Lendvay, T.S. et al., *Genetics* 25 144, 1399-1412 (1996); McEachern, M.J. & Blackburn, E.H., *Genes Dev.* 10, 1822-34 (1996)). This lethality is only observed after some delay and occurs in concert with the extensive telomere shortening observed following serial passaging of the mutant cells. Applicants reasoned that 30 mutation of other genes contributing to telomerase function, including the genes encoding the protein

- 24 -

subunit(s) of the enzyme, might be recognized because they similarly render the yeast dependent on wild-type *RAD52* for prolonged viability. If so, telomerase mutant yeast could be identified by their requirement for *RAD52* function.

5       A yeast strain carrying an inactive chromosomal *rad52Δ* allele that was complemented by a plasmid-borne wild-type *RAD52* gene linked to a *URA3* gene was generated, in order to determine if mutants in telomerase function could be revealed by a screen for mutants that require *RAD52*

10      function. When *RAD52* function was inactivated through eviction of the *RAD52*-encoding plasmid (by counter-selecting against the *URA3* marker with 5-fluoro-orotic acid (FOA), cell viability was unaffected for the succeeding 80 cell generations (Boeke, J.D. et al., *Mol. Gen. Genet.* 197, 15 345-6 (1984)). However, when the *TLC1* gene, which encodes the RNA subunit of telomerase, was additionally inactivated in this strain, a decrease in cell viability was observed after 60 generations. Moreover, when the rescuing *RAD52* plasmid was evicted, there was a marginal decrease in

20      viability after 40 generations and complete inviability after 60 generations. This showed that telomerase-deficient cells perish upon the loss of *RAD52* function and that this phenotype is therefore useful to screen for mutants carrying lesions in a variety of genes affecting

25      telomerase function.

The *rad52Δ* mutant cells carrying the complementing *RAD52* plasmid were mutagenized by homologous recombination with a library of yeast genomic fragments, each of which carried one or more copies of an inserted mini-  
30      Tn3::*lacZ*::*LEU2* transposon (Burns, N., et al., *Genes Dev.* 8, 1087-105 (1994)). To ensure a high density of mutations,

-25-

$1 \times 10^6$  transposon-mutagenized yeast clones were generated. These mutagenized yeast cells were cultured for 40 generations to allow the telomeres of any cells lacking telomerase function to shorten substantially. A portion of 5 the cell colonies was replica-plated and the *RAD52* plasmid was evicted from one of each replica pair to identify cell clones that required *RAD52* for continued viability.

Of the  $10^5$  transposon-mutagenized clones plated after 40 generations, approximately 2,500 reproducibly lost 10 viability following removal of the *RAD52* plasmid. In this case, as earlier, these cells were placed in the presence of FOA to select against the presence of the plasmid-borne *URA3* gene. The loss of viability following FOA might have derived through a second, unrelated mechanism in which the 15 *URA3* gene became stabilized through chromosomal integration, rendering cells FOA-sensitive independent of any effects on telomeres. For this reason, the FOA-sensitive cell clones were mated with wild-type yeast, yielding diploids; all cell clones for which the diploid 20 derivatives remained FOA-sensitive were discarded.

The 245 remaining *RAD52*-dependent cell clones were tested for changes in telomere structure by Southern hybridization analysis of their telomeres. Of these 245 haploids, 16 cell clones carried short telomeres identical 25 to those seen in yeast lacking a functional *TLC1* gene. Tetrad analysis of the diploid derivatives of these clones was performed to determine whether the *rad52* synthetic lethal phenotype was caused by the transposon insertion or by an adventitious unlinked mutation elsewhere in the yeast 30 genome. In each case, the lethality co-segregated with the transposon insertion, demonstrating that this phenotype is

derived directly from the genetic disruption effected by a single inserted transposon.

To determine which genes were disrupted by transposon insertion in the 16 yeast strains containing short telomeres, the transposons together with flanking genomic DNAs from each strain were cloned and the DNA sequences flanking the transposon insertion site were analyzed. This sequencing revealed that 6 clones represented 6 distinct insertion sites within the *TLC1* gene which, as mentioned above, specifies the RNA subunit of the telomerase. This result provided strong and direct validation of the utility of the strategy described to screen for genes that directly affect telomerase function.

Eight of the cell clones having shortened telomeres and lethality in the absence of *RAD52* carried transposons that had inserted into seven distinct positions within an uncharacterized open reading frame on chromosome XII at nucleotides 766540 to 769194 (Cherry, J.M. et al., Saccharomyces Genome Database (<http://genome-www.stanford.edu/Saccharomyces>)).

The predicted amino acid sequence encoded by the gene identified on chromosome XII is presented in Figures 1A and 1B (SEQ ID NO.: 2); the predicted protein includes 884 amino acids with an estimated mass of 102 kilodaltons. The predicted gene product does not share significant sequence similarity with any sequences in the databases available through the National Center for Biotechnology Information BLAST server. However, subsequent to completion of the work described herein, a report was published on the identification of a novel gene, *EST2*, involved in telomere length regulation. The open reading frame identified on chromosome XII is identical to *EST2*; therefore, this gene is referred to herein as *EST2*.

-27-

Genetic analysis of the *est2* transposon-insertion mutants shows that they have the same delayed cell death phenotype, dependence on *RAD52*, and telomere shortening as was displayed by cells carrying mutations in their *TLC1* gene. Diploids heterozygous for disrupted alleles of *RAD52* and *EST2* were sporulated and the resulting tetrads were analyzed for growth in culture and for telomere length. In 33 tetrads analyzed, the vast majority of the *est2::TN3 rad52Δ* double mutant spore products became inviable by approximately 20 generations in culture, and the remainder died by approximately 40 generations. The viability of the *est2::Tn3* mutants with wild-type *RAD52* began to decline by approximately 60 generations, and this phenotype was more pronounced after approximately 80 generations. The *EST2* wild-type spore products remained completely viable at all generations, regardless of their *RAD52* genotype. Consistent with the role of *EST2* in telomere maintenance, the decrease in telomere length, by which this gene was identified, co-segregated with the transposon insertional mutations in *EST2*. This telomere shortening was comparable to that observed in *tcl1::TN3* mutants.

Until now, *TLC1* has been the only yeast gene known to be a subunit of telomerase and to be required for telomere activity *in vitro*. The *est2::TN3* gene mutants are phenotypically indistinguishable from *tcl1::TN3* mutants. Analyses were carried out to determine whether *EST2* is also required for telomerase function.

A telomerase assay was used to show that extracts from wild-type yeast catalyze the elongation of a single-stranded telomeric primer by four to sixteen nucleotides in an RNase-sensitive fashion, indicating telomerase activity.

-28-

However, extracts derived from yeast bearing transposons inserted into *EST2* lacked detectable telomerase activity, like *tlc1Δ* mutants, which are known to be telomerase-negative (Cohn, M. & Blackburn, E.H., *Science* 269, 396-400 5 (1995)). Comparison with dilutions of wild-type extract made it possible to estimate that extracts from both *tlc1* and *est2* mutants contain at most 5% of wild-type telomerase activity. While the actual levels of telomerase in the mutant yeast are likely to be much lower, such lower levels 10 could not be quantified with this assay. As expected from the phenotypic analysis of tetrads, the absence of detectable telomerase activity co-segregated with transposon insertion, independent of the *RAD52* allele.

The requirement of *EST2* function for telomerase 15 activity suggests that Est2 is either a protein subunit of telomerase or an upstream regulator required for telomerase activity. It was possible to distinguish between these possibilities by replacing the endogenous *EST2* gene with a variant encoding a protein with three influenza 20 hemagglutinin (HA) epitope stages at its carboxy terminus (*EST2-HA*). This modified *EST2* allele and, thus, its product, is fully functional. The *EST2-HA* yeast show no overt phenotype, their telomeres are wild-type length, and extracts from these yeast exhibit normal levels of 25 telomerase activity. Extracts from *EST2-HA* and control yeast were incubated with excess anti-HA antibody, after which the immunoprecipitates and resultant supernatants were separated and assayed for telomerase activity. The anti-HA antibody immunoprecipitated telomerase activity 30 from extracts of yeast expressing the tagged Est2-HA protein, but not from extracts expressing normal Est2 protein. The immunoprecipitation was specific for the

-29-

anti-HA antibody since a control anti-Myc epitope antibody failed to immunoprecipitate activity from either extract. Essentially all of the telomerase activity remained in the supernatant of the anti-HA immunoprecipitation from wild-type extracts, whereas incubation with the anti-HA antibody almost completely immunodepleted the telomerase activity from the Est2-HA extracts. These data indicated that Est2 is not only physically associated with telomerase, but is a constituent of most, if not all, of the active telomerase complex.

Intriguingly, EST2 shares no homology with either of the two genes reported to encode the *Tetrahymena* telomerase protein subunits (Collins, K. et al., *Cell* 81, 677-86 (1995)). A search of the yeast genome (Cherry, J.M. et al., *Saccharomyces* Genome Database (<http://genome-www.stanford.edu/Saccharomyces>) revealed no open reading frame sharing obvious sequence similarity with the *Tetrahymena* genes. Thus, the genes encoding the yeast and ciliate telomerase enzymes may be highly diverged, the Est2 function may be absent in *Tetrahymena*, or the analogous Est2 protein of *Tetrahymena* may not yet have been identified.

Work described herein supports the essential role of EST2 gene for telomerase activity *in vitro* and *in vivo*. Immunochemical experiments show that Est2-containing complexes contain telomerase activity, and that removal of these complexes from the cell concomitantly removes the telomerase activity. These characteristics argue that Est2 is an essential and integral component of the yeast telomerase holoenzyme. The identification of this protein subunit of yeast telomerase should aid in the characterization of the other elements of this complex,

-30-

both in yeast and in other species, as well as in the understanding of the biochemical mechanism by which this enzyme acts.

Work described herein provides the basis for a method 5 of assessing the ability of a molecule or compound to act as a telomerase inhibitor or as a recombination inhibitor. It provides a method by which a drug (molecule or compound) which inhibits telomerase or inhibits recombination can be identified. Eukaryotic linear chromosomes are capped with 10 telomeres, a DNA-protein complex that protects the integrity of the chromosome end. Telomeric DNA sequences are composed of multiple repeat elements. When the telomeric DNA is lost from chromosome ends, cells die because of the resulting chromosomal instability.

15 Telomere length is maintained by two mechanisms. In almost all eukaryotes growing under normal conditions, telomeric DNA repeats are synthesized by the ribonucleoprotein enzyme telomerase. This RNA-dependent DNA polymerase (or "reverse transcriptase") copies the DNA 20 repeats from an RNA template that is a component of the enzyme.

When the telomerase pathway is inactive, however, telomere length can be maintained by an alternative mechanism in yeast and possibly also in humans: the 25 homologous recombination pathway used for double strand break repair.

In the yeast *Saccharomyces cerevisiae*, mutants of the telomerase subunits in conjunction with mutants of the recombination pathway have been shown to become inviable 30 after growth for 40 to 80 generations. This synthetic lethal phenotype was used as a screen for telomerase mutants in a background of recombination mutants and, as a result, a novel telomerase gene was identified.

-31-

Drug screens which work in the same fashion as demonstrated herein for the genetic screen are useful to identify telomerase inhibitors in a recombination mutant background, and recombination inhibitors in a telomerase 5 mutant background.

Telomere maintenance inhibitors, which can be a telomerase inhibitor alone, a recombination inhibitor alone, or a combination, can be identified by the method described herein, using appropriate mutant eukaryotic 10 cells, such as the mutant yeast strain described herein. Such telomerase inhibitors can be further assessed, using known methods, to confirm their effectiveness as anti-fungal drugs or as anti-cancer drugs. Normal human somatic cells lack a pathway to maintain telomere lengths, and 15 their telomeres shorten continuously. Immortal human cells, whether in tissue culture or the vast majority of malignant tumors in the human body, have developed a mechanism to maintain telomeres. Telomere maintenance occurs as a result of activation of the telomerase enzyme, 20 activation of a recombination pathway, or possibly both. In yeast, the inactivation of both pathways has been shown to kill cells; in immortal tumor cells, pharmacological inhibitors must be tested before the effects of 25 inactivation of either or both pathways can be assessed. A brief description of screens for telomerase inhibitors and recombination inhibitors follows.

#### Screen for Telomerase Inhibitors

As described below, a mutant strain of the yeast *Saccharomyces cerevisiae* that requires telomerase activity 30 for viability has been generated. This strain can be used in a rapid and automatable biological assay for telomerase inhibitors.

-32-

In the mutant yeast strain, named DN, the *RAD52* recombination gene has been replaced by a *HIS3* gene. Disruption of the *TLC1* telomerase RNA subunit gene or the *MIT1/EST2* telomerase protein subunit gene in the *rad52* 5 mutants has been shown to lead to a delayed cell death. Yeast lacking both recombination activity and telomerase activity die after about 20 to 60 cell generations.

Telomerase activity can be disrupted by enzyme inhibitors as well as by genetic inactivation, and, 10 therefore, the mutant strains described can be used to identify telomerase inhibitors. For a telomerase inhibitor screen, test compounds are added at different concentrations to the DN mutant strain and to a wild-type control strain. Each day, the cultures are be diluted 15 (e.g., 1:1000). Compounds that kill the yeast instantly, or that kill the DN and wild-type yeast with identical delays, are eliminated. Compounds that kill the DN yeast significantly before the wild-type yeast are telomerase inhibitors.

20 The method described is a biological assay and has several advantages over available methods. It is fast, automatable, and can be performed easily in large quantities. As a biological assay, it reflects the physiological function of the drugs. Furthermore, it 25 allows testing of biochemical and uptake parameters in a single step. Finally, this assay will also exclude inhibitors of other metabolic enzymes, such as RNA and DNA polymerases and is highly specific for telomerase inhibitors.

30 High-throughput screening can be accomplished by growing the yeast in microtiter dishes. Aliquoting of media, dilution of test drug compounds, and dilution of yeast cultures can be performed robotically. Growth can be

-33-

measured by the optical density of the yeast culture at 595-600 nm; growth measurements and preliminary analysis of the data can also be automated.

The telomerase inhibitor screen has several 5 applications, such as to identify inhibitors of fungal telomerase enzymes, mammalian telomerase enzymes and reverse transcriptase.

1) Inhibitors of fungal telomerase enzymes. This screen is a rapid method for identification of compounds that 10 inhibit telomerase in the yeast *Saccharomyces cerevisiae*.

This yeast is closely related to the fungal pathogen *Candida albicans* and, therefore telomerase inhibitors effective in *Saccharomyces* will likely function in *Candida*, as well as in other pathogenic yeast. There are only a few 15 effective anti-fungal drugs currently available; many are inadequate because of their serious side effects. Thus, a novel approach to anti-fungal drugs could be clinically valuable for the treatment of systemic fungal infections.

2) Inhibitors of mammalian telomerase enzymes. Telomerase 20 function is likely to be conserved between diverse species.

Therefore, it is highly possible that inhibitors of yeast telomerase could also inhibit mammalian telomerase, and can be tested for their ability to act as anti-cancer agents. These compounds can also serve as lead compounds for the 25 identification of such inhibitors and for modification to produce more effective inhibitors (e.g., inhibitors which are longer-lived, exhibit great inhibitory effects, resist degradation by cellular enzymes).

3) Inhibitors of reverse transcriptase. The drugs that 30 have been identified as telomerase inhibitors to date have been shown to be inhibitors of HIV reverse transcriptase. Telomerase inhibitors identified by the present screen can

-34-

be assessed, using known methods, for their ability to act as retroviral reverse transcriptase.

#### Screen for recombination inhibitors

In this case, inhibitors of the telomere recombination pathway are identified by their ability to kill telomerase mutant yeast but not wild-type yeast. The screen is similar to the telomerase inhibitor screen described above.

For a recombination inhibitor screen, test compounds are added at different concentrations to a telomerase mutant strain (e.g., *tlc1* or *mit1/est2*), referred to as a test strain, and to a wild-type control strain. Each day, the cultures are diluted 1:1000. Compounds that kill the yeast instantly, or that kill the telomerase mutant and wild-type yeast with identical delays, are eliminated.

Compounds that kill the telomerase mutant yeast significantly before the wild-type yeast are recombination inhibitors. The ability of the compounds to inhibit recombination can be confirmed using known methods. The recombination inhibitors have several uses. For example, combination therapy of a recombination inhibitor with a telomerase for treating fungal infections and/or malignant neoplasms. In addition, recombination inhibitors can be used to potentiate the toxic effects of radiation treatment and of chemotherapeutic agents that induce DNA double-strand breaks.

The following methods and materials were used in the work described herein.

#### METHODS

Yeast strains: Strain DN was generated by replacing the *RAD52* coding sequence of the haploid strain L3853 (gift of G. Fink; genotype *MATA leu2-3,112 lys2-201 trpl-1 ura3-*

- 35 -

52 *his3-200*) with the *HIS3* gene using a PCR-based homologous gene disruption method (Baudin, A. et al., *Nucleic Acids Res.* 21, 3329-30 (1993)). The absence of *RAD52* in this strain was complemented by introducing the 5 plasmid pYPCR<sup>+</sup><sup>35</sup>, which encodes the *RAD52* gene under its own promoter and the selectable marker *URA3*, thereby generating strain DNR. The *TLC1* gene was replaced with the *LEU2* selectable marker by the above PCR procedure in L3853 and DNR cells, yielding the strains *tlc1Δ* and *DNRtlc1Δ*, 10 respectively.

Yeast mutagenesis: Approximately 230 µg of the appropriately prepared yeast::mini-Tn3::lacZ::LEU2 genomic library (Burns, N., et al., *Genes Dev.* 8, 1087-105 (1994)) were introduced by homologous recombination into 15 approximately 7x10<sup>6</sup> cells of strain DNR. Approximately 1x10<sup>6</sup>Ura<sup>+</sup>Leu<sup>+</sup> clones representing successful transposon insertions were pooled and replated to a total number of ~1x10<sup>6</sup> clones to "age" the yeast for the *rad52Δ* synthetic lethal screen. 1x10<sup>5</sup> of such yeast were then reseeded for 20 the *rad52Δ* synthetic lethal screen, as described in the text. DNA flanking insertion sites were rescued as described by Burns, sequenced and compared to the *Saccharomyces* Genome Database (Stanford University School of Medicine, Department of Genetics) (Burns, N., et al., 25 *Genes Dev.* 8, 1087-105 (1994)).

Southern hybridization: Genomic DNA was isolated and digested with the restriction enzyme *Xba*I, which liberates the telomeric DNA (Guthrie, C., & Fink, G.R., *Guide to Yeast Genetics and Molecular Biology* (Academic Press, new 30 York, 1991); Chan, C.S. & Tye, B.K., *Cell* 33, 563-73

-36-

(1983)). 2 µg of this DNA was resolved on 0.8% agarose gel for 570 Vh, and Southern hybridized as described by Counter et al. using a <sup>32</sup>P kinase labelled yeast telomeric oligonucleotide CACCAACACCCACACACCACA (SEQ ID NO.: 37)

5 (Counter, C.M., et al., *Embo. J.* 11, 1921-9 (1992)).

Tetrad analysis: Diploid derivatives of the transposon-mutagenized yeast lacking the pYCPR+ plasmid were sporulated and tetrads were dissected according to standard methods (Guthrie, C., & Fink, G.R., *Guide to yeast genetics and molecular biology* (Academic Press, New York, 10 1991)). Presence of the rad52::HIS3 allele and the miniTn3::lacZ::LEU2 insertions were assayed on -His and -Leu synthetic complete plates, respectively.

Yeast extracts: 6 litres of yeast cultures were harvested at an optical density of 0.4-0.6 at 600 nm, resuspended in TMG buffer, disrupted with a bead beater and centrifuged for 90' at 100,000g at 4°C as described (Cohn, M. & Blackburn, E.H., *Science* 269, 396-400 (1995)). The supernatant was decanted, flash frozen and stored at ~70°C. Extracts typically had a protein concentration of 16-20 mg/ml.

Telomerase assay: Telomerase activity was assayed essentially as outlined by Cohn and Blackburn with the following two modifications: first, 4 µg of crude S100 25 yeast extract was incubated with the oligonucleotide TGTCTGGGTGTCTGGG (SEQ ID NO.: 38) and second, the reaction products were purified and resolved on 15% acrylamide, 7M urea sequencing gels (Counter, C.M. et al., *Proc. Natl. Acad. Sci. USA* 91, 2900-4 (1994)).

30 Epitope tagging: An HA-URA3-HA cassette was introduced into the 3' end of the EST2 gene by homologous recombination, and then counter-selected on FOA to excise

-37-

the *URA3* gene and generate three copies of the influenza hemagglutinin tag in frame with *EST2* (Schneider, B.L. et al., *Yeast* 11, 1265-74 (1995)). The construction of this epitope-tagged version deleted the 10 carboxy-terminal 5 amino acids of Est2; these codons contain a TA repeat sequence that is present in multiple copies in the yeast genome.

Immunoprecipitation and immunodepletion: 540 µg of yeast extract was diluted in 170 µl of TMG buffer (Cohn, M. & Blackburn, E.H., *Science* 269, 396-400 (1995)) supplemented with 150 mM NaCl (buffer TMGN) and incubated at 4°C with 20 µl of packed protein A-agarose beads pre-loaded with anti-HA antibody (monoclonal antibody 12CA5). After 2 hours, the mixture was briefly centrifuged to 15 separate the agarose beads from the supernatant extract. The beads were then washed four times with buffer TMGN and resuspended in 50 µl TMGN. As a control, a duplicate extract was diluted and left untreated on ice for 2 hours. 4 µg of supernatant or untreated extracts and 10 µl of 20 beads were assayed for telomerase activity.

#### Human Telomerase Catalytic Subunit Gene and Encoded Protein

The findings presented herein support modulation of *hEST2* (*hTERT*) RNA expression levels as an important 25 mechanism used in a variety of developmental contexts to determine the amount of telomerase activity present in specific cell lineages. Indeed, the levels of the catalytic subunit of telomerase may represent a rate-limiting determinant of enzyme activity in many types 30 of cells. Moreover, up-regulation of *hEST2* message may be an important mechanism through which telomerase becomes activated during both cellular immortalization and the progression of malignant tumors.

- 38 -

hEST2, (also referred to as hTERT) the human telomerase subunit described here, shares extensive sequence similarities with the catalytic subunits of the yeast and ciliate telomerase enzymes. The amino acid 5 sequence conservation between these three enzymes is extensive and is scattered throughout their reading frames. In addition, no related genes were detected in the human genome by Southern blotting analysis. Together, these data support the conclusion that the gene described here encodes 10 the catalytic subunit of the human telomerase holoenzyme. Further proof that this is the case can be obtained using known methods, such as experiments analogous to those performed with the yeast *EST2* gene, described herein and by Lingner and co-workers. (Lingner et al., *Science* 276:561- 15 567 (1997)). For example, experiments that will provide additional proof that the gene described herein encodes the human telomerase subunit include those that demonstrate that mice lacking the mouse *mEST2* homologue also lack telomerase activity; that the hEST2 protein is physically 20 associated with a ribonucleoprotein complex that exhibits telomerase activity; and that alteration of critical residues in the domain of hEST2 that is homologous to RTs inactivates its catalytic function.

As described herein, three telomerases described to 25 date (hEst2, yeast Est2 and Euplotes p123) include motifs that indicate they are distant homologues of a variety of RTs. They also share several unique, telomerase-specific sequence motifs. Yeast, ciliates and mammals represent highly diverged branches of the phylogenetic tree, which 30 suggests that the catalytic subunit of the telomerase was developed early in eukaryotic evolution. It is possible that it was present in the cell that became ancestral to all contemporary eukaryotes. Even earlier, it appears that

- 39 -

this enzyme shares ancestry with the precursors of the RTs specified by a variety of transposons and viruses.

The repression of *hEST2* mRNA in telomerase-negative cells and tissues and its upregulation that is associated 5 with a number of human tumor cell lines and primary human tumors suggest one mechanism by which telomerase activity might be modulated. During development, the expression of the *hEST2* mRNA may be repressed in many post-embryonic cell lineages, depriving cells in these lineages of the 10 telomerase catalytic subunit. This in turn may underlie the observed progressive telomeric shortening associated with aging in the cells in many of these lineages.

In contrast, the reappearance of telomerase enzyme activity when transformed cells escape from crisis or when 15 tumors progress toward malignancy may, in many cases, be explained mechanistically by the de-repression of *hEST2* mRNA expression. As shown here, in cell cultures, this de-repression occurs in both transformed embryonic kidney cells and lymphocytes when they emerge from crisis and 20 begin to exhibit telomerase activity. Not addressed here is whether the enhanced *hEST2* RNA expression is achieved at the transcriptional or post-transcriptional level.

Described here is a correlation between *hEST2* mRNA levels and assayable telomerase activity. These two 25 manifestations of *hEST2* gene expression are present in a constant, predictable ratio. This provides the basis for assessing cells for *hEST2* mRNA as an indicator of telomerase activity or assessing cells for telomerase activity as an indicator of *hEST2* mRNA occurrence or level. 30 It is possible that other mechanisms besides the presently observed modulation of *hEST2* mRNA levels may intervene to modulate telomerase activity.

-40-

As described in Example 8, over expression of *hEST2* in previously (normally) telomerase-negative cells is sufficient to impart telomerase activity to these cells, providing clear proof that *hEST2* is the human telomerase catalytic subunit. The data demonstrate that the ectopic expression of *hTERT* in otherwise telomerase-negative human cells is necessary and sufficient for induction of telomerase activity. In addition, the physical association of *hTERT* with telomerase activity confirms that *hTERT* is a telomerase subunit. Up-regulation of the *hTERT* gene is the sole barrier to activation of telomerase in the tested cells.

The identification of *hEST2* as the gene specifying the telomerase catalytic subunit provides an entree into understanding one of the essential steps in human tumor pathogenesis -- that leading to cell immortalization. Enzymatic assays have demonstrated that telomerase is often activated at a relatively late step in tumor progression (Harley et al., *Nature* 345:458-460 (1994)). This step may occur when evolving, pre-malignant cell clones have surmounted the senescence barrier and subsequently, having exhausted their telomeric ends, encounter the successive barrier of crisis. At this stage, the activation of telomerase may enable cells to breach these barriers to further clonal expansion, thereby conferring great selective advantage on the rare cell that has acquired the ability to resurrect the long-repressed telomerase activity. Indeed, activation of telomerase may represent an essential step in tumor progression.

Such dependence on telomerase activity means that this enzyme represents an attractive target of drugs designed to interfere with malignant cell proliferation. The finding that *hEST2* message is up-regulated in human tumors and in

-41-

immortalized cells lends further credence to this idea. Furthermore, the identification of hEST2 as the candidate telomerase catalytic subunit provides a biochemical reagent for identifying such drugs.

- 5       Described herein is a human gene, *hEST2*, which is a telomerase catalytic subunit gene. The gene transcript (mRNA) is expressed in primary human tumors, cancer cell lines and telomerase-positive tissues, but is undetectable, using the assays described herein, in cell lines known to  
10      be telomerase negative and in differentiated telomerase-negative tissues. The *hEST2* gene shares significant sequence similarity with a yeast telomerase catalytic subunit gene (*S. cerevisiae EST2*) and a ciliate telomerase catalytic subunit gene (*Euplotes p123*), as shown in Figure  
15      2. The three telomerase enzymes are members of the reverse transcriptase family of enzymes. There are seven conserved sequence motifs which define the polymerase domains of reverse transcriptases and six of these domains are present in *hEST2*, including the invariant aspartic acid residues  
20      which are required for telomerase activity. Also the subject of the present invention is DNA which encodes *hEST2* protein of SEQ ID NO.: 3; DNA which is the complement of DNA of SEQ ID NO.: 35; DNA which hybridizes to the complement of DNA of SEQ ID NO.: 35; DNA which hybridizes  
25      to the complement of DNA of SEQ ID NO.: 35 and encodes *hEST2* protein of SEQ ID NO.: 3; DNA which is a gene which localizes to human chromosome 5, subband 5p15.33. A RNA transcript or RNA message encoded by *hEST2* DNA of SEQ ID NO.: 35 or by DNA which encodes *hEST2* protein of SEQ ID  
30      NO.: 3 is also the subject of this invention.

As shown in Figure 2, conceptual translation of the 4030 bp *hEST2* DNA shows an open reading frame of 1132 amino acids which is predicted to be an approximately 127 kDa

-42-

protein. As also shown in Figure 2, there is a clear relatedness among hEST2, yeast EST2 and Euplotes p123 in regions of the gene outside the RT motifs, particularly sequence identities in a region just before motif 1 of the 5 hEST2 protein. This region of similarity appears unique to telomerases and, thus, can be used to produce reagents, such as antibodies reactive with the unique region and nucleic acid probes and primers useful to identify *hEST2* DNA, using, for example, hybridization or amplification 10 methods. *hEST2* is a single-copy gene which is approximately 40 kb in size and was shown to localize (to be present on) chromosome 5, particularly subband 5p15.33. (See Figure 4) Expression levels in normal human tissues and human cancer cell lines were assessed, as described in 15 Example 3. This assessment revealed two major RNA species (4.4 kb and 9.5 kb) and one minor RNA species (approximately 6 kb). *hEST* message was detectable in several normal tissues, including thymus, testis and intestine; the latter two tissues are known to be 20 telomerase-positive and the telomerase status of thymus has not been reported. Using the assays described herein, *hEST2* transcript was not detected in most other normal human tissues, including heart, brain, placenta, liver, skeletal muscle and prostate, all of which have been 25 reported to be telomerase-negative.

As described herein, the *hEST2* transcript or message (mRNA) is expressed in primary human tumors and cancer cell lines, but is not detectable in telomerase-negative cell lines and differentiated (normal, nontumorigenic) 30 telomerase-negative tissues. Further, *hEST2* message is not detectable in pre-crisis, telomerase-negative transformed cells, but is readily detectable in post-crisis, telomerase-positive immortalized cells. The work described

herein provides a method of differentiating between telomerase-positive cells and telomerase-negative cells, as well as for monitoring a change in telomerase content of cells. As a result, as described below, the present work 5 provides a method of identifying cells which are telomerase positive and, thus, of identifying cells which are tumor or cancer cells or are en route to becoming tumor cells (malignant).

A method of identifying telomerase-positive cells is 10 the subject of the present invention. Also the subject of the present invention is a method of identifying cells which are transformed, malignant, cancerous, tumor or post-crisis cells or are en route to or likely to become transformed, malignant, cancerous, tumor or post-crisis 15 cells. For convenience, the term malignant or malignancy is used herein to refer to transformed, cancer, cancerous, tumor, tumorigenic and post-crisis cells. For example, the phrase "a method of identifying malignant cells in an individual" encompasses a method of identifying cancer 20 cells, transformed cells, tumor cells and/or post-crisis cells in an individual. The present invention, thus, provides a method of diagnosing, aiding in the diagnosis of or predicting an increased likelihood of the occurrence of 25 cancer, tumor formation and/or the development of malignancy in an individual, particularly a mammal and, specifically, a human. In the method, telomerase content of cells is assessed by detecting or measuring *hEST2* DNA, *hEST2* transcript (*hEST2* message) or *hEST2*-encoded protein (h*EST2* protein). The quantity of *hEST2* DNA, *hEST2* 30 transcript or h*EST2* protein can be determined; alternatively, the occurrence (presence or absence) of *hEST2* DNA, *hEST2* transcript or *hEST2* protein can be detected. The presence of *hEST2* DNA, *hEST2* message and/or

-44-

*hEST2* protein in cells is indicative of the presence of malignant cells or malignancy in the individual.

In one embodiment, *hEST2* DNA, *hEST2* RNA (or both) is detected or measured in tissue or cells obtained from an 5 individual. This can be carried out using known methods, such as hybridization (e.g., *in situ* hybridization) or amplification methods. All or a portion of *hEST2* DNA or RNA can be used in such a method as a probe (e.g., to detect *hEST2* DNA or RNA) or a primer (e.g., to amplify 10 *hEST2* DNA).

As described herein, up-regulation of *hEST2* RNA is associated with activation of telomerase during cell immortalization and may be an important mechanism through which telomerase becomes activated during cellular 15 immortalization, the progression of malignant tumors or both. Thus, detection and/or measurement of *hEST2* RNA provides a means of assessing cells (e.g., mammalian, such as human cells) for malignancy or the likelihood of progression to malignancy. For example, detection of *hEST2* 20 RNA, even without quantification of *hEST2* RNA levels, in cells indicates that telomerase is activated and that the cells are malignant or have an increased likelihood of progression to malignancy (relative to cells in which *hEST2* RNA is not detected). Quantification of *hEST2* RNA can be 25 carried out as well. Higher concentrations of *hEST2* RNA cells are expected to indicate a more advanced stage in malignancy or a greater likelihood of progression to malignancy than is the case with lower *hEST2* RNA levels in cells. *hEST2* RNA levels determined by assessing an 30 individual's cells can be compared with pre-established *hEST2* RNA levels. For example, *hEST2* RNA levels determined

-45-

for an individual's cells can be compared with *hEST2* RNA levels in cells known to be malignant and/or at established stages of malignancy (e.g., a pre-established reference or standard).

5        Alternatively, the levels of *hEST2* RNA determined for an individual can be compared with *hEST2* RNA levels determined for the same individual in prior assessments, in which case the individual serves as his or her own standard or reference. In this approach, it is possible to monitor  
10      (assess over time) the status of the individual. This is useful, for example, to assess or aid in assessing whether the malignancy or cancer is progressing, remaining unchanged or regressing in that individual. This is particularly useful in assessing the status of an  
15      individual's malignancy or cancer before, during and after therapy (e.g., chemotherapy, radiation, surgery) and to assess the therapeutic value or suitability of a drug(s) or other agent(s) for a specific individual in need of treatment. As mentioned, assessment of *hEST2* RNA (or DNA)  
20      can be carried out using known methods, such as *in situ* hybridization, Rnase protection assay, reverse transcriptase (RT) PCR, Southern blot analysis or Northern blot analysis.

      In one embodiment, the presence of malignancy or an  
25      increased likelihood of malignancy is assessed in a method in which a nucleic acid (e.g., DNA) anneals to nucleic acid (e.g., RNA). For example, the method can be Northern blot analysis, Rnase protection assay or RT PCR. One embodiment is as follows: Cells to be assessed for *hEST2* DNA or RNA  
30      are obtained from the individual and processed to render DNA and/or RNA in the cells available for annealing with or hybridization of complementary nucleic acid sequences DNA or RNA (e.g., with complementary poly- or oligonucleotides

-46-

(DNA or RNA)). All or a portion of hEST2 DNA or hEST2 RNA can be used as the hybridization probe or sequence. As described herein, hEST2 protein shares sequence similarities and identities with yeast p123 and Euplotes Est2p, particularly in a region just before motif 1 (Figure 2, boxed region). As described, this region appears to be unique to telomerases. Thus, it is useful in assays which identify and/or quantify telomerase in cells and provide an assessment of malignancy or the likelihood of progression to malignancy of cells (e.g., from an individual in whom a diagnosis of malignancy or assessment of the likelihood of the occurrence of malignancy is needed). DNA or RNA comprising DNA or RNA encoding region(s) of similarity and/or identify with yeast p123 and/or Euplotes Est2p such as region(s) unique to telomerase, can be used in the hybridization assay or other assay carried out to detect or measure *hEST2* DNA or RNA in an individual's cells.

Alternatively, DNA or RNA comprising DNA or RNA which encodes a region or regions unique to hEST2 protein can be used as the probe. In an alternative approach, the *hEST2* DNA or RNA is used as a primer in an amplification method. The cells processed to render DNA and/or RNA available for annealing (hybridization) with complementary poly- or oligonucleotides (processed cells) are combined with all or a portion of *hEST2* DNA or RNA under conditions appropriate for hybridization of complementary nucleic acids (DNA, RNA) to occur. Whether annealing (hybridization) occurs is then determined. If hybridization occurs (forming complexes of cellular DNA or RNA and the *hEST2* poly- or oligonucleotides), it is an indication that the cells contain *hEST2* DNA or *hEST2* RNA. Poly- or oligonucleotides used in the method can be, for example, DNA which encodes the telomerase motif described herein (e.g., DNA which

-47-

encodes amino acid residues 556 to 565 of SEQ ID NO.: 3 or amino acid residues 560 to 565 of SEQ ID NO.: 3) or amino acid residues 1 to 50 of hEST2 protein or a portion thereof. If *hEST2* RNA is upregulated (expressed), it is 5 indicative of activation of telomerase and of malignancy or an increased likelihood of progression to malignancy. The amount of *hEST2* RNA can also be determined (by determining, for example, the extent to which hybridization with *hEST2* DNA or RNA occurs) and used to assess the extent of 10 telomerase activity and the stage of malignancy. This detection and/or measurement of *hEST2* DNA or RNA can be carried out at various intervals over time to assess the status of the malignancy (e.g., progression, reversal).

In another embodiment of assessing telomerase content 15 and, thus, malignancy or cancer of cells, hEST2 protein is analyzed (detected and/or quantified). This can be done using known methods, such as enzymatic assays or immunoassays, which indicate hEST2 protein is present in cells assessed and, optionally, quantify the protein. For 20 example, hEST2 telomerase activity can be assessed by determining whether extension of a telomeric primer occurs when a sample (e.g., cells, cell fractions or cell component(s)) from an individual is combined with the telomeric primer under conditions appropriate for hEST2 25 protein telomerase activity and telomeric extension. Alternatively, antibodies which recognize (bind) hEST2 protein can be used to determine if the protein is present in cells or other sample obtained from an individual. In the method, cells are obtained from an individual and 30 processed or treated to render proteins in the cells available for binding with antibodies, thus producing processed cells. The processed cells are combined or contacted with antibodies which bind hEST2 protein and

-48-

whether binding occurs between antibodies which bind hEST2 protein (hEST2 protein-binding antibodies) and protein in the cells is determined. If hEST2 protein-binding antibody/protein binding occurs (to form complexes), it is 5 indicative of the presence of hEST2 protein in the cells and, thus, of malignancy or an increased likelihood of progression to malignancy in the individual (in cells of the individual). The antibodies can be monoclonal antibodies or polyclonal antibodies (e.g., polyclonal sera) 10 and can be specific for (bind or recognize only) hEST2 protein or can be nonspecific for hEST2 protein (bind or recognize hEST2 protein and other protein(s)). Humanized antibodies can also be used. Antibodies which recognize a motif or epitope unique to hEST2 and/or to members of the 15 class to which hEST2 belongs are particularly useful. For example, antibodies which specifically bind the telomerase motif, represented in Figure 2, which is common to hEST2 protein, yeast p123 protein and/or Euplotes EST2 can be produced, using known methods, and used to assess cells for 20 hEST2 protein. Alternatively, antibodies which specifically bind a motif which is unique to hEST2 protein (e.g., all or a portion of amino acid residues 1 to 50 of hEST2 protein) can be used. As discussed above with reference to *hEST2* DNA and *hEST2* RNA, the assessment can be 25 detection (determination of presence or absence) or measurement (quantification). The presence of hEST2 protein is indicative of malignancy or at least of an increased likelihood of progression to malignancy.

Antibodies which bind or recognize (specifically or 30 non-specifically) hEST2 protein are also a subject of the present invention. Nucleic acid probes and primers (poly- or oligonucleotides) comprising *hEST2* DNA or RNA are also the subject of this invention. The poly- or oligonucleotides will vary in length and need to be of

-49-

- sufficient length to bind to and remain bound to *hEST2* DNA or *hEST2* RNA under the conditions used. They will generally be at least four to six bases in length and can comprise the entire *hEST2* DNA or *hEST2* RNA (alone or with
- 5 additional non *hEST2* DNA or RNA). Preferably the probes or primers will hybridize to at least a characteristic portion of *hEST2* DNA or *hEST2* RNA (a portion which is present in members of the class to which *hEST2* belongs), thus making it possible to identify *hEST2* DNA or *hEST2* RNA
- 10 substantially to the exclusion of other proteins. For example, a probe or primer can comprise DNA or RNA which encodes a characteristic motif or region of *hEST2* protein, such as the telomerase motif (e.g., amino acids 556 to 565 or 560 to 565) or the amino terminal amino acid sequence of
- 15 *hEST2* protein not present in *Est2p* or *p123*. (See Figure 2). For example, a probe or primer encoding amino acid residues 1 to 50 (inclusive) of *hEST2* protein (SEQ ID NO.: 3) or a portion thereof can be used.

Methods of altering *hEST2* DNA transcription and expression, methods of altering *hEST2* protein function and methods of identifying agents which alter (enhance or reduce) transcription, expression or function are also the subject of this invention. Also the subject of the present invention are a method of increasing or shortening the lifespan of cells in culture, *ex vivo* or *in vivo*; agents or drugs (DNA, RNA, drugs, small organic molecules, enzymes, for example) useful for lengthening or shortening cell lifespan and methods of identifying agents which enhance lifespan. For example, one embodiment of a method of

20 inhibiting *hEST2* protein comprises introducing into cells an agent which inhibits *hEST2* protein, directly or indirectly; as a result, function of the enzyme is

25

30

-50-

abolished or the enzyme is inactivated. hEST2 protein is inhibited directly, for example, by introducing into cells an agent which binds to or otherwise "ties up" hEST2 protein such that it is less active or inactive (cellular 5 telomerase activity is reduced or eliminated). Alternatively, a drug or agent which inactivates hEST2 or inhibits hEST2 catalytic function (and concomitantly reduces or eliminates telomerase activity) by degrading it or preventing it from being produced can be introduced into 10 cells in which hEST2 protein is to be inhibited. Such drugs or agents can be, for example, a small organic molecule or a dominant negative protein. Agents which prevent production of *hEST2* message or its further processing (e.g., to produce DNA) or a dominant negative 15 form of the telomerase protein can also be introduced into cells to inhibit hEST2 protein. Methods and agents which inhibit hEST2 catalytic function are useful for treatment of cancer and, thus, are useful as anti-cancer therapies.

If *hEST2* DNA, *hEST2* RNA or *hEST2* protein is inhibited 20 (partially or totally) in cells, lifespan of the cells will be shortened (shorter than it would be if the DNA, RNA or protein were not inhibited). For example, if a cell is malignant or more likely to progress to malignancy because of *hEST2* function, an agent or drug which inhibits *hEST2* 25 transcription or expression or *hEST2* protein function in the cell will also shorten the lifespan of the cell because telomerase activity (and, thus, chromosomal extension or maintenance) will be inhibited. The agent is useful as a therapeutic for treating or preventing malignancy in 30 individuals in whom malignant cells or cells with increased likelihood to progress to malignancy are present. These agents include, for example, enzyme (telomerase or *hEST2* protein) inhibitors, enzymes which degrade *hEST2* protein,

hEST2 transcriptional regulators, antisense molecules (e.g., DNA, RNA, PNA) and dominant negative mutant forms of hEST2. In a method of the present invention in which lifespan of cells is altered (increased or decreased) in an individual, a drug or agent is introduced into the individual in such a manner that it enters cells of the individual in sufficient quantity to have the desired effect (increase or decrease in cell lifespan). For example, a drug which prevents production of hEST2 transcript of hEST2 protein function (directly or indirectly) is introduced into an individual, using known methods, in sufficient quantities to enter cells, such as a tumor, precancerous or cancerous cells, whose lifespan is to be decreased (e.g., made shorter than would be the case in the absence of the drug). Alternatively, a drug which enhances cell lifespan can be introduced into an individual in such a manner that it enters cells in sufficient quantity to enhance hEST2 protein expression or prolong its activity (e.g., by blocking its degradation).

The present invention is also a method of enhancing or increasing the lifespan of cells in culture, in which telomerase activity or function is enhanced, such as by introducing hEST2 DNA or hEST2 protein into the cells or by activating an endogenous hEST2 gene. For example, the lifespan of normal human or other mammalian cells can be extended (immortalized human or other mammalian cells can be produced) and the resulting cells used for therapeutic purposes (e.g., grafting of tissue (such as skin) or of organs), screening or assay methods or production of proteins or other cellular products. For example, the lifespan of epithelial cells, keratinocytes or endothelial cells can be extended by introducing hEST2 DNA into the cells, in which the hEST2 DNA is expressed or introducing

-52-

hEST2 protein into the cells. The resulting cells with longer lifespan can be transplanted into or grafted onto an individual (e.g., as skin grafts, as systems for delivery of therapeutic proteins, such as hormones and enzymes), to whom they provide therapeutic benefit. In the method of extending the lifespan of cultured cells, cells whose lifespan is to be extended are cultured under conditions appropriate for their viability and hEST2 DNA or hEST2 protein is introduced into them. The resulting cells are maintained under conditions appropriate for expression of hEST2 DNA and/or activity of hEST2 protein, with the result that the lifespan of the cells is enhanced. At an appropriate time (e.g., after sufficient cells with enhanced lifespan are available) enhanced lifespan cells (such as keratinocytes) are transplanted into an individual (e.g., as a skin graft). cells.

The present invention is illustrated by the following examples, which are not intended to be limiting in any way.

EXAMPLE 1: Cloning of hEST2 (hTERT)

20 The expressed sequence tag database (dbEST) was searched for sequences related to the yeast protein Est2p (Lendvay et al., *Genetics* 144:1399-1412 (1996)) and the Euplotes protein p123 (Lingner et al., *Science* 276:561-567 (1997)), using the program TBLASTN and the server  
25 <http://www.ncbi.nlm.nih.gov/BLAST/>. This resulted in the identification of a homologous expressed sequence tag, Genbank accession # AA281296, which derived from Soares NbHTGBC cDNA clone 712562. DNA from clone plasmid 712562 was PCR-amplified with primers HT-1 (5'  
30 -AAGTTCCCTGCACGGCTGATGAG- 3') (SEQ ID NO.: 39) and HT-5 (5'  
-TCGTAGTTGAGCACGCTGAACAG- 3') (SEQ ID NO.: 40). The

-53-

resulting 377 bp fragment was used to probe 1 ZAP phage cDNA libraries derived from the human Jurkat T-cell lymphoma (Stratagene, La Jolla, CA) and human Nalm-6 pre-B cell leukemia cell lines (Weissbach et al., *J. Biol. Chem.* 5 269:20517-20521 (1994)). A total of seven independent clones were isolated from these two libraries. Three of these cDNA clones together with plasmid 712562 were sequenced completely in both directions. The remaining clones were sequenced in specific regions. Reprobing of 10 the Jurkat library with the 5'-most 500 bp region identified from the *hEST2* cDNA clones yielded one new clone containing an insert that overlapped with already determined sequences.

Rapid amplification of cDNA ends (RACE) was performed 15 by PCR amplifying testis Marathon-ready cDNA (Clontech, Palo Alto, CA) with flanking primer AP-1 (Clontech) and *hEST2* primer R0096 (5' -CAAGAAACCCACGGTCACTCGGTCCACGCG- 3') (SEQ ID NO.: 41), and then re-amplifying with flanking primer AP-2 and *hEST2* primer R0098 (5' 20 -CAGCTCCTTCAGGCAGGACACCTGCAGGG- 3') (SEQ ID NO.: 42). The product was subcloned and sequenced in both directions. RACE was also performed on aliquoted pools of plasmid DNA from a human testis cDNA library (Reduced Complexity cDNA Analysis, Qingyun Liu and Fang Chen, unpublished results) 25 with primers HT-21 (5' -CAGGTGACACCACACAGAAA- 3') (SEQ ID NO.: 43) or HT-22 (5' -TTCCAAGCAGCTCCAGAAA- 3') (SEQ ID NO.: 44) and a vector primer, re-amplified with the vector primer and *hEST2* primers R0098 or R0097 (5' 30 -CCTTCGGGTCCACTAGCGTGTGGCGG) (SEQ ID NO.: 45), then purified and sequenced.

DNA sequencing reactions were performed with the AmpliTaq FS Prism ready reaction cycle sequencing kit (Perkin-Elmer/ABI) and electrophoresed on a 373 A Stretch

-54-

- ABI DNA sequencer. The resulting sequences were assembled into a contig using the program Sequencher 3.0 (Gene Codes Corporation, Ann Arbor, MI). The 1132 amino acid ORF identified in this contig was aligned with p123 and Est2p
- 5 using the Pattern-Induced Multi-sequence Alignment program version 1.4 (R. F. Smith, Baylor College of Medicine & T. F. Smith, Boston University) using the server  
<http://dot.imgen.bcm.tmc.edu:9331/multi-align/multi-align.html>, with minor modifications to the final alignment.
- 10 BLASTP searches were performed against the non-redundant protein databases using the server  
<http://www.ncbi.nlm.nih.gov/BLAST/>.

EXAMPLE 2: Identification of the *hEST2* Gene

- The sequences derived from the catalytic subunit of
- 15 the *S. cerevisiae* telomerase, Est2p, and from the *Euplotes* p123 protein (Lendvay et al., *Genetics* 144:1399-1412 (1996); Counter et al., *Proc. Natl. Acad. Sci., USA* 94:9202-9207 (1997); Lingner et al., *Science* 276:561-567 (1997)), were used to identify a human homologue (Genbank accession # AA281296) in the National Center for
- 20 Biotechnology Information expressed sequence tag database. As shown in Figure 2 (underlined sequence), the conceptual translation of this DNA sequence shows clear relatedness to both the yeast and ciliate telomerase sequences as
- 25 evidenced by identical or similar amino acid residues scattered throughout the entire length of the expressed sequence tag.

- The identified expressed sequence tag provided only a fragment of the putative human telomerase open reading frame. Therefore, a human Jurkat T-cell lymphoma and a
- 30 human Nalm-6 pre-B cell leukemia cDNA library were screened

with a probe derived from the expressed sequence tag. Seven cDNA clones were retrieved. The resulting sequence was extended further in the 5' direction by rapid amplification of cDNA ends (RACE) on human testis cDNA and 5 on an independently generated human testis cDNA library.

Assembly of the cDNA clones and RACE products, together with the clone containing the expressed sequence tag, resulted in a contiguous sequence spanning 4030 bp. Conceptual translation of this 4 kbp sequence reveals an 10 open reading frame of 1132 amino acids which is predicted to encode a 127 kDa protein (Figure 2). Although we have not identified an in-frame upstream stop codon in the sequence, the first methionine identified by RACE (Figure 2) is a candidate for the translation start site, as the 15 sequence of this putative translation initiation site, CCCGCGAUGC (SEQ ID NO.: 46), is similar to the consensus GCC(A/G)CCAUGG (SEQ ID NO.: 47) characteristic of translation initiation sites (Kozak, Cell 44:238-292 (1986)).

20 The predicted 127 kDa protein shares extensive sequence similarity with the entire sequences of the Euplotes and yeast telomerase subunits (Figure 2) and extends beyond the amino- and carboxyl-termini of these proteins. A BLAST search reveals that the probabilities of 25 these similarities occurring by chance are  $1.3 \times 10^{-18}$  and  $3 \times 10^{-13}$ , respectively. By way of comparison, the probability of similarity between the yeast and Euplotes telomerases in a protein BLAST search is  $6.9 \times 10^{-6}$ . The human gene was initially named hEST2 (human EST2 homologue) 30 to reflect its clear relationship with the yeast gene, the first of these genes to be described. The current name for the gene is hTERT, which stands for human TEtelomerase Reverse Transcriptase. EST2 was named because of the

-56-

phenotype of ever shortening telomeres caused by its mutant alleles (Lendvay et al., *Genetics* 144:1399-1412 (1996)) and was later demonstrated to encode the yeast telomerase catalytic subunit (Counter et al., *Proc. Natl. Acad. Sci.*, 5 USA 94:9202-9207 (1997); Lingner et al., *Science* 276:561-567 (1997)).

Like the yeast and ciliate telomerase proteins, hEST2 (*hTERT*) is a member of the reverse transcriptase (RT) family of enzymes (Figures 2 and 3A-3F). Seven conserved sequence motifs, which define the polymerase domains of these enzymes, are shared among the otherwise highly divergent RT family (Poch et al., *EMBO J.* 8:3867-3874 (1989); Xiong and Eickbush, *EMBO J.* 9:3353-3362 (1990)). p123 and Est2p share six of these motifs with, most prominently, the a2-Sc enzyme, an RT that is encoded within the second intron of the yeast *COX1* gene (Kennell et al., *Cell* 73:133-146 (1993)). These six motifs, including the invariant aspartic acid residues known to be required for telomerase enzymatic function (Counter et al., *Proc. Natl. Acad. Sci.*, 10 USA 94:9202-9207 (1997); Lingner et al., *Science* 276:561-567 (1997)), are found at the appropriate positions of the predicted sequence of hEST2 (Figures 2 and 3A-3F). Thus, 15 the proposed human telomerase catalytic subunit, like its yeast and ciliate counterparts, belongs to the RT superfamily of enzymes.

Although hEST2 shares some sequence similarity with RTs, it is not a conventional RT. Rather, it is far more closely related to the telomerase catalytic subunits of yeast and ciliates than to other RTs. Whereas the BLAST 20 probability of sequence similarity between hEST2 and the telomerase subunits of the single-cell eukaryotes arising by chance is  $10^{-13}$  to  $10^{-18}$ , the chance probability of

-57-

sequence similarity with the next most closely related RT, a2-Sc, is 0.12. Beyond the motifs that define the polymerase domains of these various enzymes, hEST2 shows no sequence similarity with RTs. In contrast, in its domains 5 that lie N-terminal to the polymerase domain, BLAST searches identify clear relatedness between hEST2 and both p123 and Est2p, the chance occurrence of these similarities being  $1.6 \times 10^{-9}$  and  $1.8 \times 10^{-4}$  respectively. Many of the sequence identities in the N-termini of hEST2, p123 and 10 Est2p reside in a region just before motif 1 (Figure 2, boxed region). This sequence is not found in RTs, nor is it apparent in other proteins, suggesting the presence of motifs that may be unique to telomerases. Identification of the catalytic subunits of yet other telomerases will be 15 required to validate this possibility.

Even within the hEST2 domain that share sequence similarity with RTs, it is clear that hEST2 is more closely related to the already described telomerases than it is to non-telomerase RTs . For example, the sequence similarity 20 of the region encompassing the RT motifs between hEST2 and the catalytic subunits of yeast and Euplotes has a probability of chance occurrence of  $5.7 \times 10^{-6}$  and  $1.9 \times 10^{-5}$  respectively compared to 0.0056 for a2-Sc, the next most closely related non-telomerase RT. Within the RT 25 motifs are several amino acids that are invariant among the telomerases but divergent between telomerases and non-telomerase RTs, or alternatively nearly invariant among non-telomerase RTs but divergent between these RTs and telomerases (Figures 3A-3F ).

30 In summary, hEST2, Est2p, and p123 form a clearly defined subgroup within the RT family. For these reasons, hEST2 is a human homologue and very likely an orthologue of the microbial enzymes described to date.

-58-

Sequencing of a number of cDNA clones has revealed two distinct forms of *hEST2* transcripts. See Figures 5A-5B and Figures 7A-7C. Four independent cDNA clones, isolated from three independently generated libraries deriving from 5 distinct cell types, lack an identical 182 bp segment within the open reading frame. The absence of this segment leads to a shift in reading frame that introduces a premature termination codon. Both forms of the *hEST2* transcript were detected by RT-PCR in a variety of human 10 cell types. Information on the intron-exon boundaries of *hEST2* is not available. The simplest interpretation of these data is that the sequence difference between the two groups of cDNAs reflects the existence of two alternatively spliced mRNAs of the *hEST2* gene. The physiological 15 consequences of the expression of the potential non-functional *hEST2* transcript are obscure at present.

**EXAMPLE 3: Chromosomal Localization of *hEST2***

The *hEST2* cDNA was used as a probe in Southern blot analyses of human genomic DNA. These reveal *hEST2* to be a 20 single-copy gene with an estimated size of 40 kb. All the genomic sequences reactive with the cDNA probe appear to be components of this ~40 kb locus, suggesting that there are no other closely related genes in the human genome. *hEST2* was localized to a specific chromosomal region 25 by analyzing two independent panels of hamster-human radiation hybrid (RH) cells with two markers spanning different regions of *hEST2*. Initial mapping using the Genebridge 4 RH panel placed both *hEST2* markers between sequence-tagged sites (STS) WI-9907 and D5S417 (Figure 4). 30 Independent confirmation of this localization was then obtained by mapping carried out with a second panel, the

-59-

Stanford G3 RH panel. This second mapping placed both *hEST2* markers next to STS marker AFMA139YA9 (GDB locus D5S678) which itself is localized between the above-mentioned markers WI-9907 and D5S417. These markers 5 are present at the telomeric end of chromosome 5p (Hudson et al., *Science* 270:1945-1954 (1995)).

Mapping of the *hEST2* locus was further refined by localizing it to subband 5p15.33, since the STS markers D5S678 and D5S417 have been assigned to this band on the 10 Genethon YAC contig map (Chumakov et al., *Nature* 377:175-297 (1995)). This localization is consistent with fluorescence in situ hybridization (FISH) analysis of YAC 767E1. This YAC maps further away from the telomere than D5S678 and has been assigned to chromosome 5p15.33 by FISH 15 (Chumakov et al., *Nature* 377:175-297 (1995)).

Chromosome 5p is one of the most common targets for amplification in non-small-cell lung cancers; it is amplified in ~70% of tumors (Balsara et al., *Cancer Res.* 56:645-650 (1997); Petersen et al., *Cancer Res.* 57:2331-2335 (1997)). The effects of this amplification on *hEST2* expression levels are unknown.

EXAMPLE 4: Expression of *hEST2* Transcripts

Telomerase becomes activated during tumor progression. and, as discussed above, this activation has been 25 associated with the immortalization of tumor cells. A variety of mechanisms might be invoked to explain such activation, among which is the induction of the expression of one or more telomerase subunits. The telomerase holoenzyme is presumed to exist as a multi-subunit 30 ribonucleoprotein complex and, therefore, the levels of any one of the subunits, including those of the catalytic

-60-

subunit described here, might be rate-limiting in determining enzyme activity. Alternatively, the components of the telomerase holoenzyme might be expressed constitutively and subject to various types of 5 post-translational modification that would govern their activity. As mentioned earlier, transcript levels of the telomerase RNA subunit and of the *TP1/TLP1* gene do not necessarily correlate with telomerase activity (Feng et al., *Science* 269:1236-1241 (1995); Avilion et al., *Cancer Res.* 56:645-650 (1996); Blasco et al., *Nat. Genet.* 12:200-204 (1996); Harrington et al., *Science* 275:973-977 (1997); 10 Nakayama et al., *Cell* 88:875-884 (1997)), implying that at least one other mechanism is responsible for regulating human telomerase.

15 The expression levels of *hEST2* mRNA in various cell types were analyzed, using both RNA Northern hybridizations and RNase protection assays, in order to address one of the remaining possible regulatory mechanisms,. RNA blots prepared from a panel of normal human tissues and human 20 cancer cell lines were probed with cDNA fragments deriving from two independent, non-overlapping regions of the *hEST2* gene. This probing revealed two major RNA species migrating near the 4.4 kb and the 9.5 kb markers, as well as a minor species of ~6 kb (Figures 8A-8F). Each of these 25 RNA species was recognized by both probes, confirming that each represents an *hEST2* mRNA.

*hEST2* message was detectable in several normal tissues including thymus, testis, and intestine. Of these, testis (Kim et al., *Science* 266:2011-2015 (1994); Wright et al., 30 *Dev. Genet.* 18:173-179 (1996)) and intestine (Hiyama et al., *Int. J. Oncol.* 9:453-458 (1996)) are known to be

-61-

telomerase-positive, while the telomerase status of the thymus has not been reported. In marked contrast, the *hEST2* transcript was undetectable in our assays in most other normal human tissues, including heart, brain,  
5 placenta, liver, skeletal muscle, and prostate (Figures 8A 8B, 8D and 8E), all of which have been reported to be telomerase-negative (Kim et al., *Science* 266:2011-2015 (1994); Wright et al., *Dev. Genet.* 18:173-179 (1996); Shay and Bacchetti, *Eur. J. Cancer* 33:787-791 (1997)). The  
10 absence of detectable *hEST2* message in ovary may seem paradoxical, as ovary is a germline tissue, and germline tissues have been reported to harbor significant levels of telomerase. However, oocyte division is completed by birth, and while fetal ovary is telomerase-positive (Kim et  
15 al., *Science* 266:2011-2015 (1994); Wright et al., *Dev. Genet.* 18:173-179 (1996)), both adult ovarian epithelium and mature oocytes are telomerase-negative (Counter et al., *Proc. Natl. Acad. Sci., USA* 91:2900-2904 (1994); Wright et al., *Dev. Genet.* 18:173-179 (1996)). It is also possible  
20 that the levels of *hEST2* are below the threshold of detection. The 7 kb band seen in muscle tissue appears to be an artifact, as it was not observed with an independent *hEST2* probe.

In contrast to its absence in the majority of normal  
25 tissues, *hEST2* mRNA was strongly expressed in a variety of cancer cell lines, most strikingly in the leukemic cell lines HL-60 and K-562 (Figures 8C and 8F). It is unclear why only very low levels of *hEST2* message were observed in HeLa cells on this particular Northern blot (Figures 8C and  
30 8F); reanalysis of independently prepared HeLa cell RNA both by Northern blot and by RNase protection demonstrated

- 62 -

that *hEST2* mRNA is present in HeLa cells at high levels comparable to those seen in K-562 cells. The two major and one minor *hEST2* transcript appear to be expressed in the same relative proportions in all cell types that yielded 5 detectable *hEST2* mRNA.

EXAMPLE 5: *hEST2* Expression in Primary Human Cancers

The expression of *hEST2* mRNA in cancer cell lines suggested that *hEST2* transcript levels might be elevated in primary tumors as well. To determine if this is the case, 10 RNA was extracted from a variety of tumor samples as well as from normal control tissues and analyzed this for the presence of *hEST2* mRNA using an RNase protection assay. In total, 11 of 11 tumor samples examined showed detectable levels of *hEST2* message (Figures 9A-9D). *hEST2* RNA was 15 undetectable in normal breast and ovarian tissue but was expressed at significant levels in 2 of 2 breast tumors as well as 2 of 2 breast tumor-derived cell lines, and in 4 of 4 ovarian tumors.

As expected from Northern hybridization analysis of 20 *hEST2* RNA (Figures 8A-8F) and the known pattern of telomerase catalytic activity in human tissues (Kim et al., *Science* 266:2011-2015 (1994); Hiyama et al., *Int. J. Oncol.* 9:453-458 (1996)), the *hEST2* transcript was detected at high levels by the RNase protection assay in testis and at 25 moderate levels in colon. Similarly, using the RNase protection assay, four colon tumor samples and a testicular tumor sample were found to express detectable levels of *hEST2* RNA (Figures 9A-9D); two of the colon tumors showed significantly elevated levels as well. These data suggest 30 that the *hEST2* message is expressed in a very high

- 63 -

percentage of tumors and is specifically induced in tumors that arise from telomerase-negative tissues.

EXAMPLE 6: Up-regulation of *hEST2* mRNA Is Associated with Telomerase Activation and Cellular Immortalization

5       The strong expression of the *hEST2* message observed in several tumors and cancer cell lines suggested that the levels of this transcript are correlated with the amount of telomerase enzyme activity in these various cell types. To investigate this possibility, telomerase activity and *hEST2* RNA level were analyzed in a panel of non-immortalized and immortalized cell lines. Using the TRAP telomerase assay 10 (Kim et al., *Science* 266:2011-2015 (1994)), two mortal fibroblast strains, WI-38 and IMR-90, were found to lack detectable telomerase activity. In contrast, telomerase 15 activity was readily detectable in three immortal cell lines, HeLa, 293, and K-562. The telomerase-negative cells also lacked detectable *hEST2* message as gauged by an RNase protection assay, while the immortal telomerase-positive cell lines expressed significant levels of *hEST2* RNA. 20 These results indicate that *hEST2* message levels correlate closely with telomerase activity.

Thus, as described herein, *hEST2* RNA expression and telomerase activity are present in immortal transformed cells, but absent in mortal normal cells. These findings 25 support the role of induction of *hEST2* expression in the activation of telomerase that occurs during cellular immortalization. That this is the case was shown by analyzing *hEST2* transcript levels by RNase protection, and comparing the levels of hTR and telomerase activity in 30 pre-crisis cells prior to the up-regulation of telomerase, and in post-crisis telomerase-positive immortal cells from

-64-

- two different transformed human cell populations: Epstein-Barr virus-transformed B lymphocytes (B4 cells) and SV40-T antigen transformed embryonic kidney cells (HA1 cells). Result observed were consistent with previously 5 reported assessments (Counter et al., *EMBO J.* 11:1921-1929 (1992), *Proc. Natl. Acad. Sci., USA* 91:2900-2904 (1994); Avilion et al., *Cancer Res.* 56:645-650 (1996)). Little or no telomerase activity was detected in cells prior to crisis when telomere length decreases, but abundant levels 10 were detected in post-crisis cells which maintain telomere length. In contrast, the levels of *hTR* remained essentially constant throughout the immortalization process. The levels of *hEST2* RNA, on the other hand, parallel telomerase activity during cell immortalization. 15 *hEST2* RNA was undetectable in the pre-crisis cells but was clearly present in the post-crisis, telomerase-positive cells. Induction of *hEST2* message is the rate-limiting step for the activation of telomerase during immortalization.
- 20 EXAMPLE 7: Down-regulation of *hEST2* Expression upon Cellular Differentiation
- The up-regulation of *hEST2* RNA is associated with the activation of telomerase during cell immortalization. It remained unclear whether conditions that repress telomerase 25 activity in cultured cells might similarly operate by shutting down expression of the *hEST2* mRNA in these cells. Human HL-60 promyelocytic leukemia cells can be induced to differentiate to mature granulocytes by treatment with retinoic acid. During this process, telomerase activity 30 has been shown to decline over a period of two to five days (Sharma et al., *Proc. Natl. Acad. Sci., USA* 92:12343-12346

-65-

(1995); Bestilny et al., *Cancer Res.* 56:3796-3802 (1996); Savoysky et al., *Biochem. Biophys. Res. Commun.* 226:329-334 (1996); Xu et al., *Leukemia* 10:1354-1357 (1996)).

Induction of HL-60 differentiation was shown to lead  
5 to the disappearance of the *hEST2* mRNA within 24 hours,  
foreshadowing the loss of telomerase activity which  
declines more slowly. Once again, this contrasts with the  
behavior of the human telomerase RNA subunit, whose  
expression during induced differentiation remains constant,  
10 as also previously reported (Bestilny et al., *Cancer Res.*  
56:3796-3802 (1996)). Results presented here show that the  
levels of the *hEST2* mRNA decline precipitously within the  
first three hours after induced differentiation. This  
rapid down-modulation of is compatible with a short  
15 half-life for the *hEST2* message and suggests that the  
levels of this RNA species are under tight control in these  
cells. The contrasting delay in the decline of telomerase  
activity is consistent with the reported ~24 hour  
half-life of the enzymatic activity after cycloheximide  
20 treatment (Holt et al., *Mol. Cell. Biol.* 16:2932-2939  
(1996)).

EXAMPLE 8 Ectopic Expression of hTERT mRNA in Telomerase-Negative Cells and Assessment of Expression on Levels of Telomerase Activity

25 The *hTERT* cDNA was introduced into a mammalian expression construct carrying the CMV promoter. To distinguish the ectopically expressed *hTERT* protein from its endogenous counterpart, the C-terminus of the vector-encoded protein was marked with an influenza virus  
30 hemagglutinin (HA) epitope tag, yielding the plasmid

-66-

pCI-neo-*hTERT-HA*. Such a modification does not affect the catalytic activity of the Est2p protein of *S. cerevisiae* (Counter, C.M., et al., "The catalytic subunit of yeast telomerase", *Proc. Natl. Acad. Sci. USA*, 94:9202-9207 5 (1997)).

This *hTERT-HA* construct and a control empty vector were transfected into cells of the SV40-transformed GM847 human fibroblast line (Pereira-Smith, O.M., and Smith, J.R., "Genetic analysis of indefinite division in human 10 cells: identification of four complementation groups", *Proc. Natl. Acad. Sci. USA*, 85:6042-6046 (1988). In contrast to most immortal human cells, which appear to activate telomerase in order to maintain telomere length (Shay, J.W., and Bacchetti, S., "A survey of telomerase 15 activity in human cancer", *Eur. J. Cancer*, 33:787-791 (1997), the immortal GM847 cells are telomerase-negative (Bryan, T.M., et al., "Telomere elongation in immortal human cells without detectable telomerase activity", *EMBO J.*, 14:4240-4248 (1995). Moreover, while the *hTR* gene is 20 transcribed in these cells (Bryan, T.M., et al., "The telomere lengthening mechanism in telomerase-negative immortal human cells does not involve the telomerase RNA subunit", *Hum. Mol. Genet.*, 6:921-926 (1997), they lack detectable levels of *hTERT* mRNA (Kilian, A., et al., 25 "Isolation of a candidate human telomerase catalytic subunit gene, which reveals complex splicing patterns in different cell types", *Hum. Mol. Genet.*, 6:2011-2019 (1997). These observations have led to the conclusion that GM847 cells employ a telomerase-independent mechanism to 30 maintain telomere length (Murnane, J.P., et al., "Telomere dynamics in an immortal human cell line", *EMBO J.*,

-67-

13:4953-4962 (1994); (Bryan, T.M., et al., "Telomere  
elongation in immortal human cells without detectable  
telomerase activity", *EMBO J.*, 14:4240-4248 (1995), perhaps  
analogous to the recombination-based pathway used by yeast  
5 cells to allow chromosomes to maintain telomeres in the  
absence of a functional telomerase pathway (Lundblad, V.,  
and Blackburn, E.H., "An alternative pathway for yeast  
telomere maintenance rescues est1- senescence. *Cell* 73,  
347-360 (1993). Unlike normal, telomerase-negative human  
10 somatic cells, which lack replicative immortality, these  
GM847 cells could be propagated indefinitely following  
transfection, allowing us to study the properties of  
clonally isolated cell populations that have stably  
acquired the introduced *hTERT* gene.

15 A number of stably transfected GM847 cell clones were  
generated with either the control vector or the *hTERT-HA*  
expression vector, mRNA expression of *hTERT* was analyzed by  
RNase protection using probes that specifically recognize  
either the transfected *hTERT-HA* mRNA or the endogenous  
20 *hTERT* transcript. As expected, *hTERT-HA* transcript was  
detected only in GM847 sublines stably transfected with an  
*hTERT-HA* expression plasmid, but not in untransfected  
telomerase-positive control cell lines 293 and HL-60, in  
the parental GM847 line or in GM847 sublines transfected  
25 with the empty vector. The cells expressing *hTERT-HA* did  
not express the endogenous *hTERT* transcript, despite the  
fact that this RNA is clearly detected in telomerase-  
positive cells. Lastly, in accord with previous  
observations that the level of the *hTR* RNA subunit of  
30 telomerase does not correlate with enzyme activity, this  
RNA was detected in all cells tested, irrespective of  
whether the cells had telomerase activity. An actin

-68-

control probe demonstrates comparable loading of RNA from each cell line. Moreover, the specificity of the probes used was demonstrated by their failure to protect tRNA.

Expression of hTERT-HA was also analysed at the 5 protein level, by immunoblotting with an anti-HA antibody probe directed against the HA tag of the vector-encoded hTERT protein. A ~ 130 kDa product corresponding to the predicted size of hTERT was detected in those lines derived from GM847 cells that were stably transfected with the 10 hTERT-HA expression construct, but not in those cell clones that had been transfected with the empty vector. The antibody likewise did not detect endogenous (untagged) hTERT known to be expressed in 293 cells (Meyerson, M., et al. "hEST2, the Putative Human Telomerase Catalytic Subunit 15 Gene, Is Up-Regulated in Tumor Cells and during Immortalization," *Cell*, 90:785-795 (1997), a telomerase-positive control cell line.

Assessment of whether these hitherto telomerase-negative GM847 cell lines acquired telomerase activity 20 together with the stable ectopic expression of hTERT was carried out. Telomerase activity was measured in these different cell lines by assaying the ability of a cellular extract to elongate a primer in a telomerase-specific manner. The products of this *in vitro* reaction are 25 subsequently detected by specific PCR amplification, yielding a ladder of products differing from one another by 6 bp (Kim, N.W., et al., "Specific association of human telomerase activity with immortal cells and cancer", *Science*, 266:2011-2015 (1994)). Telomerase activity was 30 detected in 293 cells, a cell line with one of the highest levels of telomerase activity known. This activity was sensitive to heat treatment of the extract, which inactivates telomerase (Kim, N.W., et al., "Specific

-69-

association of human telomerase activity with immortal cells and cancer", *Science*, 266:2011-2015 (1994)). In contrast, almost no telomerase products were detectable following assay of extracts from untransfected GM847 cells 5 or from GM847 sublines stably transfected with the empty control vector. This inability to detect telomerase was not due to the presence of a PCR-inhibiting activity, as an internal control was specifically PCR-amplified, (Kim, N.W., and Wu, F. "Advances in quantification and 10 characterization of telomerase activity by the telomeric repeat amplification protocol (TRAP)", *Nucleic Acids Res.*, 25: 2595-2597 (1997).

In marked contrast, telomerase activity was readily detectable in those clones of GM847 cells that were stably 15 transfected with the *hTERT-HA* expression vector and this activity was sensitive to heat inactivation. The levels of telomerase activity observed in these transfectants approached those seen in extracts from 293 cells. The restoration of telomerase activity in the cells transfected 20 with *hTERT-HA* was not due to the up-regulation of the endogenous *hTERT* gene, as evidenced by failure to detect the corresponding mRNA in these cells. Thus, ectopic expression of *hTERT* in previously telomerase-negative cells 25 is sufficient to generate telomerase activity at levels comparable to those found in immortalized telomerase-positive cells.

The telomerase activity detected in cells transfected with the *hTERT-HA* expression vector was physically associated with ectopically produced *hTERT-HA*, confirming 30 that *hTERT* is, as predicted, a constituent of the telomerase holoenzyme. Telomerase activity could be immunoprecipitated with an anti-HA monoclonal antibody from

-70-

extracts of either 293 cells or GM847 cells that ectopically express *hTERT-HA*. The telomerase activity was not immunoprecipitated with an antibody directed against an irrelevant antigen (anti-p53), nor was it

5 immunoprecipitated when either antibody was incubated with extracts prepared from control vector-transfected GM847 cells. Taken together, these data indicate that telomerase activity is specifically co-immunoprecipitated with *hTERT-HA*.

10 Whether the observed induction of telomerase activity following ectopic *hTERT* expression was unique to GM847 cells was also determined. This was done by transiently transfecting WI-38 normal human fibroblasts, which lack detectable levels of telomerase activity and *hTERT* message  
15 but express the hTR gene (Meyerson, M., et al., "hEST2, the Putative Human Telomerase Catalytic Subunit Gene, Is Up-Regulated in Tumor Cells and during Immortalization. Cell 90, 785-795 (1997), with the *hTERT-HA* expression construct. Since transient transfection of normal human  
20 cells is extremely inefficient, the plasmid pGreenLantern-1, which encodes the green fluorescent protein (GFP), was co-transfected with either the *hTERT-HA* expression vector or the control vector. Cells expressing GFP, and hence quite likely the co-transfected plasmid,  
25 were sorted by virtue of their fluorescence.

Extracts derived from both populations of fluorescing cells were assayed for telomerase activity. Whereas fibroblasts transfected with pGreenLantern-1 and pCI-neo vector alone lacked enzymatic activity, those  
30 co-transfected with the *hTERT-HA* expression vector were clearly telomerase-positive. Transfection with pCI-neo-*hTERT-HA* of IMR-90 cells, another telomerase-

- 71 -

negative normal human fibroblast cell strain that does not normally express hTERT (Kim, N.W., et al., "Specific association of human telomerase activity with immortal cells and cancer", *Science*, 266: 2011-2015 (1994);  
5 (Meyerson, M., et al., "hEST2, the Putative Human Telomerase Catalytic Subunit Gene, Is Up-Regulated in Tumor Cells and during Immortalization", *Cell*, 90: 785-795 (1997), also gave rise to telomerase activity (not shown). Thus,  
ectopic expression of hTERT in these two types of normal  
10 human somatic cells results in readily detectable telomerase activity.

Taken together, these data demonstrate that the ectopic expression of hTERT in otherwise telomerase-negative human cells is both necessary and sufficient for induction of telomerase activity.  
15 Furthermore, the physical association of hTERT with telomerase activity confirms that hTERT is a telomerase subunit. The fact that forced expression of hTERT sufficed to impart telomerase activity indicates that levels of hTR mRNA and TP-1 or other still unidentified components of the telomerase holoenzyme are not rate-limiting determinants of telomerase activity in these cells. Up-regulation of the hTERT gene is therefore the sole barrier to activation of telomerase in the tested cells.  
20

25 It still remains to be determined if telomerase activity can be restored in this fashion in all telomerase-negative cells. However, results described here show that activity is conferred by ectopic expression of hTERT in cell types representative of two known classes of telomerase-negative cells: telomerase-negative immortal  
30 cell lines and normal mortal human cell strains. Thus, it is likely that expression of hTERT mRNA is the

- 72 -

rate-determining step for telomerase activation in other human cells lacking enzyme activity. This makes the regulation of transcription from the *hTERT* promoter a potential target for modulation during tumorigenesis and  
5 cell immortalization.

#### Experimental Procedures

The following methods and materials were used in the experiments which are described above.

#### Radiation Hybrid Mapping

10 25 ng of genomic DNA from hybrid clones of the Genebridge 4 and Stanford G3 radiation hybrid (RH) mapping panels (Research Genetics, Inc. Huntsville, AL) was PCR-amplified with the primer pair M1 (forward-5'  
-CACAGCCAGGCCGAGAGCAGA- 3' (SEQ ID NO.: 48) and reverse-5'  
-AGGCCTCAGCCGGACACTCAG- 3') (SEQ ID NO.: 49), yielding a 170  
15 bp fragment in the 3'-untranslated region, and with primer pair M2 (forward-5' -GAAGAAAACATTTCTGTCGTG- 3' (SEQ ID NO.: 50) and reverse- 5' -GCCCTTGGCCCCCAGCGACAT- 3') (SEQ ID NO.: 51), generating a 180 bp fragment crossing a putative  
20 intron-exon boundary near the *hEST2* stop codon. PCR was carried out for 35 cycles of 94°C for 0.5 min., 69°C for M1 or 65°C for M2 for 0.5 min. and 72°C for 1.5 min. The results of the PCR screening were analyzed using the statistical program RHMAP provided through the following  
25 two e-mail servers on the World Wide Web:  
<http://shgc-www.stanford.edu> and  
<http://www-genome.wi.mit.edu/cgi-bin/contig/rhmapper.pl>.  
Linkage of STS markers to a physical chromosomal map was achieved by accessing the servers:  
30 <http://www.cephb.fr/ceph-genethon-map.html> and  
<http://www-genome.wi.mit.edu>. Comparison to YACs

- 73 -

physically mapped by FISH was achieved with the server:  
ftp://ftp.cephb.fr/pub/ceph-genethon-map/FISH/29MAR95.DAT.

#### RNA Isolation from Cell Lines and Primary Tissues

Cell lines were obtained from the American Type  
5 Culture Collection and grown under standard conditions.  
For HL-60 differentiation assays, cells were pelleted and  
resuspended in normal growth medium plus all-trans retinoic  
acid (Sigma) at a final concentration of 1  $\mu$ M.  
All primary normal and tumor tissues were obtained  
10 from the Massachusetts General Hospital tumor bank.  
Tissues were processed in a tissue homogenizer. RNA  
samples were prepared in the RNA Stat-60 solution (Tel-Test  
"B", Friendswood, Texas) according to the manufacturer's  
protocol.

#### 15 Northern Hybridization

Duplicate filters containing poly A(+) -selected mRNAs  
from various human tissues and cell lines (Multiple Tissue  
Northern Blots, Clontech, Palo Alto, CA), were incubated  
according to the manufacturer's instructions with two  
20 independent *hEST2* probes. One probe was derived by  
PCR-amplifying plasmid 712562 with primers HT-1 and HT-5 to  
generate the 377 bp fragment described above; the second,  
more 3' probe was generated from the 1064 bp *Stu* I fragment  
of the same plasmid. The duplicate blots were then  
25 rehybridized with a  $\beta$ -actin probe. Northern blots for  
detection of *hTR* were performed with total RNA and an  
*hTR*-specific probe (Feng et al., *Science* 269:1236-1241  
(1995)).

#### RNAse Protection Analysis

30 Radiolabelled RNA probes for RNAse protection analysis  
were synthesized using [ $\alpha$ -<sup>32</sup>P]UTP, T7 RNA polymerase and

-74-

the MAXIscript kit (Ambion, Austin, TX). DNA templates for probe synthesis were created as follows. For the *hEST2* probe, the insert from plasmid clone 712562 was subcloned into pUHD 10-3 (a gift from M. Gossen), and template DNA 5 was synthesized by PCR using a forward plasmid-specific primer and a reverse primer containing 18bp of *hEST2* complementary sequence (5'-TCTCTGCGGAAGTTCTG) (SEQ ID NO.: 52) and the T7 promoter sequence. For the  $\beta$ -actin probe, template DNA was synthesized by PCR on a human  $\beta$ -actin cDNA 10 insert (Clontech). The *hEST2* and the  $\beta$ -actin control probes were hybridized in the same reaction tube. The *hTR* probe was synthesized directly from the linearized pGRN83 plasmid (Feng et al., *Science* 269:1236-1241 (1995)).

RNase protection analysis was performed using the 15 HybSpeed RPA kit (Ambion) according to the manufacturer's protocol. Briefly, sample RNA and radiolabelled RNA probes were coprecipitated in ethanol, resuspended in hybridization buffer, hybridized at 68° C, then digested with RNases A and T1. Samples were then re-precipitated 20 and analyzed on a 6% denaturing polyacrylamide gel.

#### Telomerase assays

Telomerase repeat amplification protocol (TRAP) assays were performed as described (Kim et al., *Science* 266:2011-2015 (1994)), in some cases using the TRAPeze telomerase 25 detection kit (Oncor, Gaithersburg, MD).

#### EQUIVALENTS

While this invention has been particularly shown and described with references to preferred embodiments thereof, it will be understood by those skilled in the art that 30 various changes in form and details may be made therein without departing from the spirit and scope of the

-75-

invention as defined by the appended claims. Those skilled in the art will recognize or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described  
5 specifically herein. Such equivalents are intended to be encompassed in the scope of the claims.

-76-

CLAIMS

What is claimed is:

1. Isolated DNA encoding the catalytic subunit of a eukaryotic telomerase holoenzyme.
- 5 2. Isolated DNA of Claim 1 which encodes the catalytic subunit of a yeast telomerase holoenzyme.
3. Isolated DNA of Claim 2 wherein the DNA is selected from the group consisting of: DNA of SEQ ID NO.: 1; DNA which hybridizes to the complement of SEQ ID NO.: 1; and DNA which encodes the amino acid sequence of 10 SEQ ID NO.: 2.
4. Isolated DNA of Claim 1 which encodes the catalytic subunit of a human telomerase holoenzyme.
5. Isolated DNA of Claim 4 wherein the DNA is selected 15 from the group consisting of: DNA comprising the nucleotide sequence of SEQ ID NO.: 35; DNA which encodes hEST2 protein having the amino acid sequence of SEQ ID NO.: 3; and DNA comprising a nucleotide sequence which hybridizes to the complement of SEQ ID 20 NO.: 35.
6. Isolated DNA of Claim 4 which is an isolated gene which localizes to human chromosome subband 5p15.33.
7. Isolated DNA comprising DNA encoding a human 25 telomerase catalytic subunit, wherein the DNA encodes an RNA transcript which is up-regulated in a human tumor.

- 77 -

8. Isolated DNA of Claim 7 which is an isolated human gene which localizes to human chromosome subband 5p15.33.
9. Isolated DNA of Claim 7 which comprises *hEST2* cDNA of SEQ ID NO.: 35 or DNA which hybridizes to the complement of *hEST2* cDNA under conditions of high stringency.
10. Isolated DNA of Claim 7 wherein the human tumor is selected from the group consisting of: breast tumors, ovarian tumors, colon tumors and testicular tumors.
11. Isolated DNA of Claim 7 which encodes a human telomerase catalytic subunit which comprises approximately 1132 amino acid residues.
12. Isolated DNA of Claim 8 wherein the RNA transcript is also up-regulated in a cancer cell line, a telomerase-positive tissue or both.
13. Isolated mRNA which encodes a human telomerase catalytic subunit, wherein expression of the mRNA is up-regulated in a human tumor.
- 20 14. Isolated mRNA of Claim 13 wherein the human tumor is selected from the group consisting of: breast tumors, ovarian tumors, colon tumors and testicular tumors.
15. Isolated mRNA of Claim 13 wherein the mRNA has the sequence of SEQ ID NO.: 36.

-78-

16. A nucleic acid probe comprising DNA unique to *hEST2* DNA, *EST2* DNA and *p123* DNA.
17. The nucleic acid probe of Claim 16 comprising DNA which encodes the telomerase motif of *hEST2* protein.
- 5 18. The nucleic acid probe of Claim 17 comprising nucleotides which encode amino acid residues 556 to 565 of SEQ ID NO.: 3 or nucleotides which encode amino acid residues 1 to 50 of *hEST2* protein or a portion thereof.
- 10 19. A method of assessing cells for malignancy or an increased likelihood of progression to malignancy, comprising:
  - a) obtaining cells to be assessed from an individual;
  - 15 b) processing cells obtained in a) to render DNA and/or RNA in the cells available for annealing of the DNA and/or RNA with complementary poly- or oligonucleotides, thereby producing processed cells;
  - 20 c) combining processed cells with *hEST2* DNA or RNA under conditions appropriate for annealing of DNA and/or RNA with complementary poly- or oligonucleotides, thereby producing a combination,
- 25 wherein if annealing occurs in the combination produced in c), it is indicative of activation of telomerase and malignancy or increased likelihood of progression to malignancy.
20. The method of Claim 21 wherein the poly- or  
30 oligonucleotides are DNA which encodes the telomerase

- 79 -

motif or amino acid residues 1 to 50 of SEQ ID NO.: 3 or a portion thereof.

21. A method of diagnosing or aiding in the diagnosis of development of malignancy in an individual,  
5 comprising:
  - a) obtaining cells from the individual;
  - b) processing the cells obtained in a) to render proteins in the cells available for binding with antibodies;
  - 10 c) combining the product of b) with antibodies which bind hEST2 protein; and
  - d) determining whether binding occurs between the antibodies which bind hEST2 protein and protein in the cell,
- 15 wherein if binding occurs, it is indicative of the presence of hEST2 protein in the cells and of malignancy or an increased likelihood of development of malignancy in the individual.
22. The method of Claim 21 wherein the antibodies which bind hEST2 protein bind the telomerase motif of hEST2 protein or bind amino acid residues 1 to 50 of SEQ ID NO.: 3 or a portion thereof.  
20
23. A method of reducing expression of *hEST2* RNA and hEST2 protein in cells of an individual, comprising  
25 administering to the individual a drug selected from the group consisting of: drugs which inhibit or bind *hEST2* RNA and prevent or reduce production of hEST2 protein and drugs which inhibit hEST2 protein function or activity.

24. The method of Claim 23 wherein the drug is selected from the group consisting of: small organic molecules; enzymes which degrade hEST2 protein; enzyme inhibitors; HEST2 transcriptional regulators; antisense molecules and dominant negative forms of hEST2 protein.
- 5  
25. A method of treating cancer in an individual, comprising administering to the individual a drug which inhibits or binds *hEST2* RNA and prevents or reduces production of hEST2 protein or a drug which inhibits hEST2 protein function or activity, under conditions appropriate for the drug to enter cells of the individual, whereby the drug inhibits or binds *hEST2* RNA and prevents or reduces production of hEST2 protein or inhibits hEST2 protein function or activity.
- 10  
15  
26. The method of Claim 25 wherein the drug is selected from the group consisting of: enzymes which degrade hEST2 protein; enzyme inhibitors; hEST2 transcriptional regulators; antisense molecules and dominant negative forms of hEST2 protein.
- 20  
27. A method of increasing lifespan of cells, comprising:
  - (a) introducing into the cells *hEST2* DNA, under conditions appropriate for expression of *hEST2* DNA, whereby *hEST2* protein is expressed or
  - 25  
(b) introducing *hEST2* protein into the cells, whereby sufficient *hEST2* protein is expressed or present in the cells to increase lifespan of the cells.

- 81 -

28. The method of Claim 27, wherein the cells are cells in culture.
29. A method of decreasing lifespan of cells in an individual by reducing hEST2 protein function or activity, comprising administering to the individual a drug selected from the group consisting of:
  - (a) drugs which inhibit or bind *hEST2* RNA and prevent or reduce production of *hEST2* protein;
  - (b) drugs which inhibit *hEST2* protein function or activity; and
  - (c) drugs which inhibit or bind *hEST2* DNA and prevent or reduce production of *hEST2* RNA transcript, wherein the drug is administered to the individual under conditions appropriate for entry of the drug into cells in sufficient quantity to reduce *hEST2* protein function or activity.
30. The method of Claim 29 wherein the drug is selected from the group consisting of: small organic molecules; enzymes which degrade *hEST2* protein; enzyme inhibitors; *HEST2* transcriptional regulators; antisense molecules and dominant negative forms of *hEST2* protein.

1 / 14

ATGAAATCTTATTGGAGTCATTCAAGACAAGTTCATTGATCTACAGGACAACTTACAGGACTTAAAGAATTAAATGTTGGTCACCTCAATGGCC  
 M W K I L F Q D K L D I D L Q T N S T Y K E N L K C G H F N G  
 TCGATGAAATCTAACTACGGTTTCGCACTACCAATTCAAGAAAATGGCATTACCATGCCTTCGTTGACTTAAGCCACAAAGCAGTCATTGATCA  
 L L D B I L T C F A L P N S R K I A L P C L P G D L S H K A V I D H  
 CTGCAATTACTGTGCTTAAATGCTGAGAATGTTGCTTAACTACGTTACATAGCATTGGCTTAACTGCTAACATAAGTCTTGGC  
 C I I Y L L T G E L Y N N V L T G Q F F T Q I V G N R C N E P H L P P K W V Q R S S S  
 CATTCTGCAATGTTAACGTTACGTTACTGTAAGGGCGCTGCTGTTAACAGGTTAACAGATGTTACGAACCTCATCGGCCCAAATGGGTCAACGATCATCCT  
 H S A N V N V T L L K G A A W K M F H S L V G T Y A F V D L L I N  
 ATACAGTAAATTCAATTCAATTGCGCTGTTTCACTCAATTCTACACAGCTAACAAATTCAAGGCTAACATAATTCTCTTC  
 V I Q F N G Q F P V T N K Q F L H K L N I N S S S F F P  
 ATCATCGCAACTGGCTGCGCAATTCAACAACTTACAGAACCTGAGAACTTACGTTGAGAAGCTTACCTCCACAAATTGGTTAAATTCTCAGAGACTAA  
 S S A T A A Q I K O L T E P V T N K Q F L H K L N I N S I L N S I C P P L E G T  
 TATAGCAAGATCCCTCTTCATCATCTACAAAGCTTAACCTGAGAAGCTTACCTCCACAAATTGGTTAAATTCTCAGAGACTAA  
 Y S K I L P S S I K K L T D L R E A I F P T N L V K I P Q R L  
 AGGTACGAATTGTTGACGCTGCAAAAGCTTAAAGGACATAAGGGTTGAAATTACGTTCTTAAATGGCTTAAATGTTGAGAAGCTTACGTTGCTC  
 K V R I N L T L Q K L L K R H K R L N Y V S I L P L N G Y L P F D S L L K K  
 CGTATGGACTGTCGCATTGAGTAGGCATTCAAAAGGAACGAGCTGTTGAIAATTATTCTAGGTTTACAGAATTACGGCAATTGGCGATTGGTTC  
 V L D L S H L S R Q S P K B R V L L Q P K H N F B N L N Q L A I C F I S W  
 GGCTCAAAAGAAAAATTATCAAGATCTAAATCTTATTAGGTTACCCCTTAAGGAAACAAACGTTGAAAMCCAATAGTTACAGGTTGCTC  
 G S K K N K G K I I P F Y C T E I S S T V T I V Y F R H D T W  
 TAAGATTAAAGGATTTCGGTTGTTCACTTGTTGATTTGTTGATATTGGTTCACTTACAGCTTAAATGTTGAGGATTATCA  
 L R L S D I W F T K H N F B N L N Q L A I C F I S W  
 GCTATTAGACAACTAATTCCCAAAATTATACAGACTTTTACTGCACCGGAAATTACTCTACAGTCAATTGTTACTTACGTTAGACATGTTGCG  
 L F R Q L I P K I I Q T F Y C T E I S S T V T I V Y F R H D T W  
 AATAAACCTTACCCCTTTATCGTGAATTATTAGAGCTTAACTTACAGCTTAAATGTTGAGGATTATTCGTTGATTTGTTGAGGATTATCA  
 N K L I T P F I V E Y F K T Y L V E N N V C R N H N S T L S N F  
 ATCATAGGAAATTAGGATTATACCAAAAAAAAGTTAAATGAGTTAACTACAGACTTTTACTGAAACAAACGACTTAAAGGAAATTTTA  
 N H S K M R I I P K K S N N E F R I I A F P C R G A D E E F T I Y  
 TAAGGAGAAATCACAATTGCTTAAAGGATTATCGGCCACTAAAAATTAGAATACCTAAGGAAACAAAGGCGACTAGTTTACTAAATTATTC  
 K E N H K N A I Q P T Q K K I L E Y L R N K R P T S F T K I Y S P T  
 CAAATAGGTTGACGGTCAAGGAAATTAGGAGACTTAAAGGAAATTAGGTTGAGGACTTAAAGGAAATTTTAATAGTGTGTTCAATTCTC  
 Q I A D R I K E F K Q R L L K K D A L K N E N G F F V R S Q Y F F N T  
 GCTATTAGGTTCAAGGATGCGAAATTGTTGAGGATACTCAGGATGCGCTAAAAAATGAAATTGTTGAGGTTGATCTCAATTCTCAATTAC  
 C Y D S I P R M E C M R I L K D A L K N E N G F F V R T V H L S  
 CAATACAGGTTATTGAAGGTTATTATGTTGAGGTTAACGCTAGGACGTTAACGTTAACATAGATAATTGTTGAGGACGGTTCAATTCTC  
 N T G V L K L F N V V N A S R V P K P Y E L Y I D N V R T V H L S  
 AATCAGGATGTTATAAACGTTGAGGATGGCAAAATTAAACGCTTGTGCTACATTAGGAAGGATAAGTGTGCTTCAATTGGGTTGAGGCT  
 N Q D V I N V V E M E I F K T A L W V E D K C Y I R E D G L F Q G

FIG. 1A

2/14

CTTAGTTTATCTGGTCCGATCGTGTGATTGGTGTATGACGATCTTCTGGAGTTTATAGCGAGTTAAGCCAGTCCTAGCCAGGACCATTAATTTPAAA  
 S S L S A P I V D L V Y D D L E F Y S P K A S P Q D T L I L K  
 ACTGGCTGAGGATTTCCTTATAATATCAACAGACCAACAGCAAGTGTATCATATCAAAAGCTTGCCTGGCATTGCCAATGGGATTTCABAATAATGCGAAGGCC  
 L A D D F L I S T D Q O Q V I N I K K L A M G F Q K Y N A K A  
 AATAGAGACAAATTAGCCGTAAGCTCCCAATCGATGATGATACGGTTTATTCATGGCAATGGCAATATTGTTAAAGAATTGGAAAGTTGGAA  
 N R D K I L A V S S Q S D D T V I Q F C A M H I F V K E L E V W  
 AACATTCAGCACCATGAAATTCCATATCCGTAAGGGATATTTCGAAGGTTAATAGCGCTGTTAACACTAGAAATCTCTTATAAA  
 K H S S T M N N F H I R S K S K G I F R S L I A L F N T R I S Y K  
 AACATTGACACAAATTAAATTCAACAAACACCGTTCTCATGCAAATTGATCATGTTGAAGAACATTGGAAATGTGTATAAAATCTGCTTTAAGGAT  
 T I D T N L N S T N T V L M Q I D H V V K N I S E C Y K S A F K D  
 CTATCAAATTAAATGTTACGCAAATATGCAATTTCATTGTTCAACGCATCATGAAATGACAGTCAGGGTTGTCCAATTAGAAATGTGATCCTT  
 L S I N V T Q N M Q F H S F L Q R I I E M T V S G C P I T K C D P  
 TAATCGAGTATGGGTTACGATTCCACCATATTGAATGGATTGGAAAGCCTATCTCTCAACACATCAAATTAAAGATAATATCATTCTTTGAGAAA  
 L I E Y E V R F T I L N G F L E S L S S N T S K F K D N I I L L R K  
 GGAAATTCAACACTTGCAAGCATAATATAACATATAATGTTAATTAG  
 E I Q H L Q A Y I Y I H I V N  
 2655

FIG. 1B

3/14

M P R A P R C R A V R S L L S H Y R E V L P L A T F V R R L L G P Q G W R L V Q O G D P A A F R A L H E S T Z  
 51 V A Q C L V C V P W D A R P P P A P S F R Q V S C L K E R S V A R Y L O R L C E R G A K N V L A F G F A I B L D G A R G G H E S T Z  
 1 M E V D V O N Q A D N H G I H S A L K T C E I K A T O Y S - W I T Q K V I - R C R N S Q S H Y K D E D I K I F A P 123  
 1 M K I I F E - F I Q D K L D I D L Q T N S T Y K E N K C K G H F N G I E S T Z P  
 1111 P P E A F F T E S V N S L P N T V T D A L R G S G A W G L L B L R R - V G D D O V E T H E A R C A L F V L V A P S C A Y H E S T Z  
 59 Q T N T V A T P - R D V N E E D F K V I A R K E V F S T G E M I E - L I D K C L V E L U S S - - - S D V S D R Q K L Q P 123  
 34 L D E I L T T - - - C F A L P N S R K I A L P C L P G D I S H K A V I D H C I Y L L - - - - - T G E L Y N N E S T Z P  
 170 N C S P P L Y Q G A A T Q A R P - P P H S G P R R R L G C E R A N N H S W P E A G V P L G L P A P G A R R R G G S A H E S T Z  
 113 C F G - - - F O R K G N Q L A K T H L L T A L S T Q K Q Y F F Q D E W N - Q V R - - - - - A M I G N E R F R H L P 123  
 81 M L T T - - - F E S Y K I A R - - - N E D V V N S L - - - - - F E S T Z P  
 229 S R S L P L P K E P R R G A A P E P E R T P V G Q G S W A H P G C T R G P S D R G F C V V S P A R P A E E T S L E G A H E S T Z  
 160 Y T K Y L I F O R T S E G T L V Q - - - F C G N N V F D H A L K V N D K F D O K K - - - - - O K G G A N D M N E P R P 123  
 100 C H S A N V N V N T L L K G A A W K M F H S L V G T Y A F V D I L L I N Y T V I Q F N - - - Q G F F T Q I V G N R C N E P H E S T Z P  
 289 D S G F R H S H P S V G R Q H H A G P P S T S R P R P D T P C P P V Y A E K H E L Y S S S G D K - E Q L R P S F L L H E S T Z  
 208 C C S T C K Y N V K N E K D H F - - - L N N I N V E N - W N N - - - M K S R F R I Y C T H F N R N N Q F F K K H E F P 123  
 157 L P P K W V Q R S S S S A T A A - Q I K Q L T E D - V - - - - - T N K Q F L H K L N I N S S S F P Y S K I E S T Z P  
 348 S S L R P S L T G S P L R V E T I F L G S R P W M P G T P K R R L P R P P Q R Y W Q M R P L F L L L G N H A Q C P X G V H E S T Z  
 260 Y S N K N N I S A M D R - A Q T I F - - T N I F R F N I R K - - K D K V I E K I A Y M E E K V D F - - - N F N Y P 123  
 205 L P S S S S K K L L D R E A I E - P T N L V K I P Q - - - R L K V R I N L T L Q K L U K R H K R L N - - - X V S E S T Z P  
 408 L W K T H D R A A V T A A G V C A R E P Q Q G S V A A P E E E D T D P P R R L V Q L I D R Q H S S P W Q W Y G - E V R H E S T Z  
 312 Y T K S C P L - - - B E N W R E R K Q K I E N L I N K T R E E K S - - - K Y Y E E N F S Y T T D N W K C V T Q F E T N P 123  
 256 I N S I C P - - - P L E G T V D - - - - - L S H I L S R Q S P K E R V Y L - - - K F E I E S T Z P  
 467 A C R R V V P P G L W G S R H I N E R R L R N T K F I S I G K H A K L S V R G C A W R R S P G V H E S T Z  
 364 E F F Y N I L P K D F L W G T R K N F Q K K V K V K K Y V E L N K H L U L L E K I N T R E I S M Q V E T S A P 123  
 289 V R I Q K L L P Q E M F G S K K N K G K I I K N L N L L S I P L N G Y L P F D S L L K K K L R K D F R W L F I - - S D E S T Z P  
 527 G C P A A E H R L R E E I A K F L H W L M S V Y V E C L L R S F F Y V T E T T F Q K N R K F F Y R K S V W S K L Q S H E S T Z  
 423 K H E Y Y F D H E - N I V E W K L R W I V E S T C F E Y U T E Q Q S Y S K T K Y V K N I W D I W K P 123  
 347 I W E T K H N F E N N O L A C E F I S W F R O T P K I T O F F Y C T E I S S T V - T I V V F R H D T W N K S I T E S T Z P

FIG. 2A

4 / 14

**motif 1**  
 587 I G E R Q H T R V Q I R E L S E A E V R Q H E A R -- P A L L T S R I R F I P K - - P D G L R P I - - V N M D Y - V V hEST2  
 482 M S E - A D U - K E T - A E V Q E K E V E W K K S - - L G D A P G K L R L I P K - - K T T F R P I - - M T F N - - - p123  
 486 P F F - V E Y F K T Y V - - E N N V C R N H S Y T L S N P N H S C K M R I I P K K S N N E F E R T I A I P C R G A D E Est2p  
  
**motif 2**  
 641 G A R T F R R E K R A E R I E T S R V C A N F V N Y E R A R R P G L - - L G A S T L G L D D I T H R A W R T F E A L R V hEST2  
 531 - K K I V N S D R K T T K T T N T L N S H E M L K T L K N R M F K O P F S A V F N Y D D V M K K Y E E F V C C N p123  
 462 E E F T I Y K E N H K N A I Q P T Q I T - - E Y T R N K E P T S F T - - K I Y S P T Q I A - D R I T K E F K Q R L L K F F Est2p  
  
**motif 3**  
 698 R A T D P P P E L Y F V R D V T G A Y D T H Q D R I T E V I - - - - -  
 590 K - - V G Q P K L F F A T M D I E K C Y D S V N R E K U S T F E K T T K L L S S D F W I M T A Q I T K R K N N I V I D S p123  
 518 - - N N V L P E L Y M R F D V K S C Y D S I P R M E C W R I L K - - - - -  
  
**motif 4**  
 744 Y A V V Q A A H G H V S K A E K S H V S T T T D L S - - P Y M R Q F A H L D E T S P L R D A V V T E Q S S S L N E A S hEST2  
 649 K N T R K V E M K O Y F N Q K N Q K - - I A E G G Q Y E T I L H S V L E N E Q D L N A K K T T I V E A K Q R N Y F K K p123  
 562 Q Y F N T N T - - - - - G V L - - - - - K L F N V  
  
**motif 4**  
 803 S G A F D W F L R F N C H H A V R I R G K S Y V O C O G I T P O G S I I S T I L C S L C Y G D M E N K L F A G I T R D G - hEST2  
 707 D N L Q P V I N I C Q N Y I N F N G K F Y K O T K G C P Q G L C V S S I L S S F Y Y D V A L E S S L G F L R D E S M p123  
 602 Q D V I N W A E M E I F K T A L W V E D K C Y I R E D G L F O G S S S S A P I V D L V Y D D L L E - F Y S E F K A S P - Est2p  
  
**motif 5**  
 862 - - - - - L L L R L V D D E L L V I P H L T H A K L B I L R T W U R C V P E Y G C V V N L R K T V V N E P V E D E A hEST2  
 767 N P E N P N V N L L M R L T D D M L L I T P Q E N N A V L V I E K I E V E Y - - - - -  
 660 - - - S Q D T L I K K L A D D E L L I S D Q Q Q - V I N I K K L A M G - - - - - G F Q K Y N A K - - - - - A N R O K I Est2p  
  
**motif 6**  
 914 T G G T A F V Q U P A R G L - - - F P W C G I L L D I T R I T E W Q S D Y S S Y A R T S T R A S I S T F M - - R G F K M G R hEST2  
 827 F - - K Y G M D S V E E Q N I V Q D Y C D W I G I S S E D M R C T I A L N P N I - N L R I E G I L C T N L N M Q T K K p123  
 706 T A V S S Q S D D O T - - - - - V I Q F Q A M H I F V K E E Y - - - - - W K H S I S T M N N F H I R - - - S K S S K Est2p  
  
**motif 7**  
 969 N M R B E E F G V E R L K C H S L E L O L Q V N S L Q T S V C T N I Y I T L L Q A S T R F H A V L Q L P F H Q Q V W K N hEST2  
 886 W L K K K K K S F S M N N E T H Y - - R K T I T O E D F A N K T L N K L F I S G G K Y M Q Q A K E - - Y K D H F K K N p123  
 , 751 G I F R S I I A L F N T R E - - - - - S Y K I D T I O T I L N S T N T V M Q I D H V V K N I S E - - - C Y K S A F K D Est2p  
  
**motif 8**  
 1029 P T F F L R V I S O T A S L C Y S I L K A K N A G M S L G A K G A A G P L P S E A V Q W I U C H Q A F L L K L - - T R H R V hEST2  
 943 P A M S S M I D L E V S K I I Y S V T R A F F K Y L V C N I K D T I F G E E H P D F F E S T L K H F I E I F S T K K Y p123  
 801 U S I N V T Q N M Q F H S F L Q R E I V E N T V S G C P I T K - - C D P T I E V E R T I L N G F I E S R - S S N T S Est2p  
  
**motif 9**  
 1088 T Y V P L L G S I T A Q T Q - - S R K L P G T T T T A L E A A N P A L P S D F K T I L D hEST2  
 1003 T E N R V C M I K K A K E A K Q S D Q C - - Q S E I Q Y D A Q A Y I Y I V I H V  
 857 K K D N T I I L K R E I Q H Q A Y I Y I V I H V

FIG. 2B

5/14

**MOTIF 1**

hEST2	SRLR	FIPK		
p123	GKLRL	IPIK		
Est2p	SKM	RITIK		
a2-Sc	VRRVEIPIK			
a1-Sc	MRIIVNIPIK			
motif	p	hh	h	K

**MOTIF 2**

LRPTIVNMDYVVG	
FRPIMTFLNKKV	
FRIIAIPCRGAD	
FRPLLSVGNPREK	
TRPISVGNPRDK	
hR	K
R	

FIG. 3A

**MOTIF 3**

*	DVTGAYDTI
PELYEVKVD	
PKLFFATMDIEKCYDSV	
PELYFMKFDVKSCYDSI	
YCNWEIKVDLNKCFDTI	
GSNWFEREVDLKKCFDTI	
hDh	AF
GY	

FIG. 3C

**MOTIF 4**

KSYVQOCOGIPQGSILSTLLCSLQY	
KFYKOTKGIPQGLCVSSILSSFY	
KQYIREDGFLFQGSSLSSAPIVDLVY	
NYHNTTLGIPQGSVVSPILCNIFL	
TYHKPILGLPOGSLSIPILCNIVI	
hpQG	hp
hh	hh
h	h

FIG. 3D

**MOTIF 5**

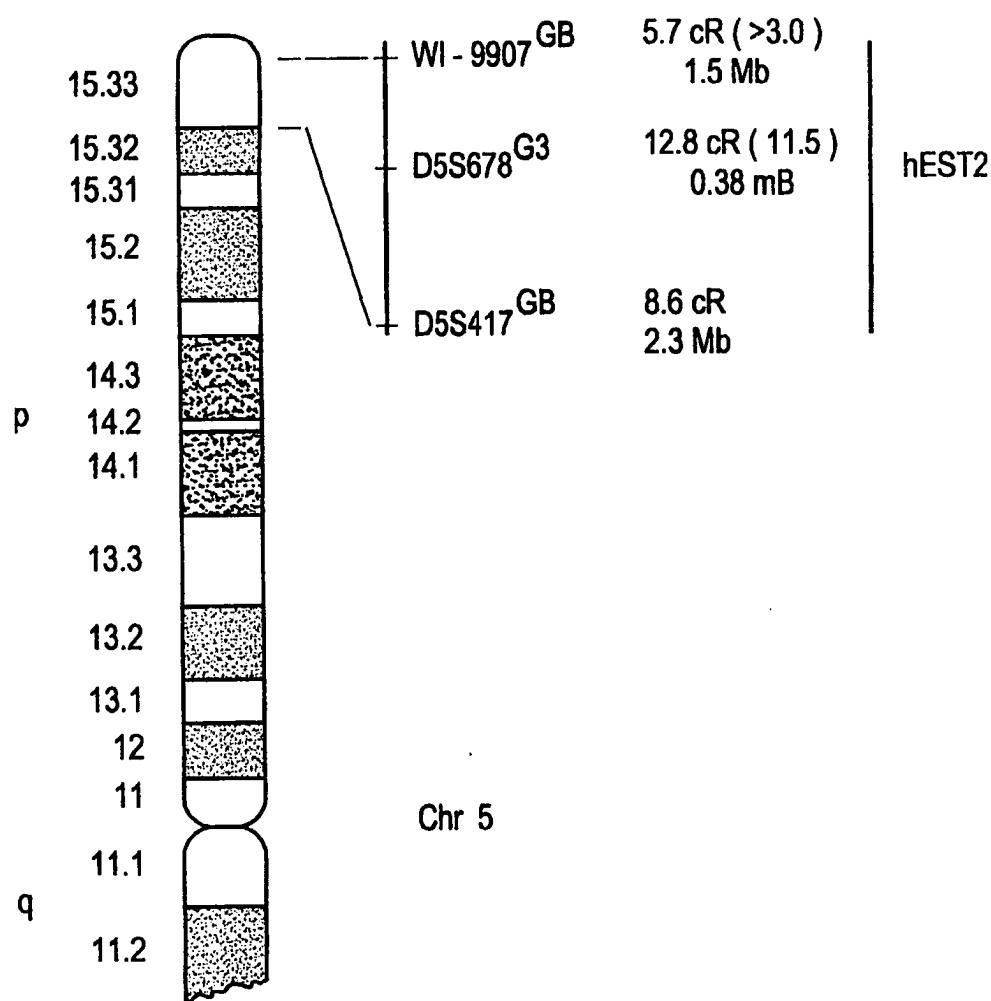
LLRLVDDFLVLT	
LMRLTDYLLIT	
LILKLAADDFLLIS	
YFVRYADDIIIGV	
KYVRYADDIIIGV	
h	Y
DDhhh	

FIG. 3E

**MOTIF 6**

GCWNLRKTVV			
GFKFNMKLLQT			
GFQKYNAK---			
GMSINIDKSVI			
GLTINEEKTLI			
Gn	h	CK	h

FIG. 3F

**FIG. 4**

7/14

	10	20	30	40	50	60	70	80
CAGGGCTGGCTGGTCTGCTGGCACGTGGAAAGCCCTGGCCCCGGCCACCCCCGGATGCCGGCTCCCCGGCTGCC	80							
GAGCCCTGGCTCCCTGCTGGCAGCCACTAACCGCAGGGTCTGCCGCTGGCCACGTTCTGTGGCCCTGGCCCAG	160							
GGCTGGGGCTGGCTGGTCAAGGG	240							
GGCACGG	320							
TGTGGGAAGGG	400							
TTTCAACCACGGTGGCGAGCTACCTGGCCAAACAGGGTACCCGAGCGCACTGGGGGGGGGGGGGGGGGGGGGG	410	420	430	440	450	460	470	480
GCGCCGGCTGG	490							
ACCAGGGTGTGGATGGGATGGGAAACGGGGCTGGAAACCATAGGGTAGGGGGGGGGGGGGGGGGGGGGGGGG	560							
AGGGGTCTGGGATGGGATGGGAAACGGGGCTGGAAACCATAGGGTAGGGGGGGGGGGGGGGGGGGGGGGGGGG	640							
GAGGG	720							
CCAGGG	800							
GGACGCCGTTGGGGCAGGGGTCCTGGGGCCACCCCCGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	810	820	830	840	850	860	870	880
CCCTGGCAGACCCGGCGAAGGCCACCTTGTGG	880							
CCAGGACACTGG	960							
CCAGGACACTGG	1040							
CCAGGACACTGG	1120							
ACTGGGGCTGGGGGGGGCTGGGAGGACCATCTTCTGGGGTCCAGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	1200							
CCTGGCCCCAGGGCTACTGGCAATGGGGCCCTGGTCTGGGAGCTGGTCTGGGAGCTGGCTGGGGGGGGGGGG	1210	1220	1230	1240	1250	1260	1270	1280
TCTCTCAAAGAGCCACTGGCCGGCTGGGAGCTGGCTGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	1280							
GTGG	1360							
GTACGGCTTCTGG	1440							
TCAGGAAACAAAAGGTTCACTCCCTGG	1520							
CGGCTTCACTCCCCAGGG	1600							
1610	1620	1630	1640	1650	1660	1670	1680	
GGGGGCTGGCTGGGGCCAGGG	1680							
GGCCAAAGTTCTGCACTGGCTGATGAGGTGGTACGTGGCTGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	1760							
TTCAAAAGAACGGGCTTCTGGGGAGGTGGAAAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	1840							
AGGGTGAGCTGGGGAGGTGGAAAGGG	1920							
CCGCTTCACTCCCCAGGG	2000							
2010	2020	2030	2040	2050	2060	2070	2080	
AAAGAGGG	2080							
CTCCCTGGGGGGGGCTTGTGGCTGG	2160							
CCCGGG	2240							
TCATCGGGCAAGCACTCAAAACCCCAAGAACAGGTACTGGGTGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	2320							
GTTCGG	2400							

FIG. 5A

8/14

	2410	2420	2430	2440	2450	2460	2470	2480
GAGACCAAGCCCCGGCTGAGGGATGCCGTGTCATCGAGCAGAGCTCTCCCTGAATGAGGCCAGGAGTGGCCAGTGGCTCTTGACG	2490							
TCTTCCATACGCTTCATGTGCCAACACGCCCTGGCATCAGGGGAAAGTCTTAAGTCCACTGTCAGTCCAGTGGCCAGGGGATCCCGCAGGGC	2560							
TCCATCCTCCACGGCTGTGCTCTCACGGTGGATGATTCTGGATGATTCTGGTTGACACCTCACCCACGGAAACAGCTGGTGAACCTGG	2640							
GCTGGCTCTCGCTTGGTGAACACTGGTGAACACTGGCTGTCGAGTGGCTGAGTGGCTGAGTGGCTGAGTGGCTGAGTGGCTGAGTGGCTG	2720							
TCCGAGGGTGTCCCTGAGTGGCTGAGTGGCTGAGTGGCTGAGTGGCTGAGTGGCTGAGTGGCTGAGTGGCTGAGTGGCTGAGTGGCTG	2800							
	2810	2820	2830	2840	2850	2860	2870	2880
GGTGGCACGGCTTTGTCAGATGGGGCCACGGCTTATCCCCTGGTGGCCAGTGGCTGCTGGATACCCGGATCCGGCTGGGA	2880							
GGTGCAGAGGACTACCTCCAGCTATGCCGGACCTTCATCAGAGGCCAGTCTACCTTCATCAACGGCTTCAAGGGCTTCAAGGGCTGGGA	2960							
GGAACATGCTGCCAAACTCTTGGGTCTTGGGGCTGAAGTGTCAAGGCCCTGGATTGGAGGTGAACAGCCTC	3040							
CAGAGCTGGCACCAACATCTACAAGATCTCCTGTCAGGGTGTGAGGGTGTGAGGGTGTGAGGTGGCTGAGTGGCTGAGTGGCTGAGTGG	3120							
TCTACATGGCAAGTGGAAACCCACATTTCCTGGCTGCGTCATCTGACGGCCCTCCCTGGTACTCCATCTGAGTGGCTGAGTGGCTGAGTGG	3200							
	3210	3220	3230	3240	3250	3260	3270	3280
AAGCCAAGAACGGAGGGATGTTGCTGGGGGCCAGGGGCCAGGGCCCTCTGCCCTCGAGGGCCGTGAGTGGCTGTCAGTGGCTGTCAG	3280							
CACCAAGACATTCTGGCTCAAGGTGACTGACCTACGTGGCTCACTGGGGGAGCAACGGCTGGGGGAGCAAGGGGGAGCACTGGGGCTCA	3360							
GGAGCTGAGTGGAAAGTCTGGCAACGGCTGGGGAGCAAGGGGGAGCAACGGGGGGAGCAAGGGGGAGCAACGGGGGGAGCAACGGGGGGAG	3440							
AGACCATCTGGACTGTGGCAACGG	3520							
GTCCCCAGGG	3600							
	3610	3620	3630	3640	3650	3660	3670	3680
CTGCATGTCGGGCTGAAGGCTGAGTGTCCGGCTGAGCGAGTGTCCAGGCCAAGGGCTGAGTGTCCAGGCCAAGGGCTGAGTGTCCAGACACCT	3680							
GGCGTCTTCACTTCCCAAGGGCTGGGCTCAGGGCCACCCCAAGGGCTTTCCTCCCTGGCCCTGCCCCATGTTCAACCCCTGCCCCATGTTCA	3760							
CCCCACATAGGAATAATGTCACATCCCAAGATTCGCCATGTTCAACCCCTGCCCCATGTTCAACCCCTGCCCCATGTTCAACCCCTGCCCCAT	3840							
TCCAGGTGGAGACCCCTGAGAAGGACCCCTGAGAAGGAGCTGGGAATTTGGAGTGGCAAAAGGTGGCTGACAGGGCTGACAGGGCTGAG	3920							
GACCCTGGCACCTGGATGGGGCTCCCTGTTGGCTCAATTGGGGGAGGGTGTGGGAGTAAATACTGAAATATGAGTTGAGT	4000							
	4010	4020	4030	4040	4050	4060	4070	4080

FIG. 5B

9/14

M P R A P R C R A V R S L L R S H Y R E V L P L A T E V R R L G P Q G W R L V Q R G D P A A F R A L V A Q C L V C V P W D A R P P P A P S F R  
Q V S C L K E L

V A R V I L Q R L C E R G A K N V L A F G F A L I D G A R G G P P E A F T T S V R S Y L P N T V T D A L R G S G A W G L I I L R R V G D D V L V H  
L I L A R C A L F V

L V A P S C A Y Q V C G P P L Y Q L G A A T Q A R P P H A S G P R R R L G C E R A W N H S V R E A G V P L G L P A P G A R R G G S A S R S L

P L P K R P R R  
R P W D T P

G A A P E P E R T P V G Q G S W A H P G R T R G P S D R G F C V V S P A R P A E E A T S L E G A L S G G T R H S H P S V G R O H H A G G P S T S R P P

C P P V Y A B T K H F L Y S S G D K E Q L R P S P L L S S L R P S L T G A R R L V E T I F L G S R P W M P G T P R R L P R L P Q R Y W Q M R P L F L  
E L L G N H

A Q C P Y G V U L K T H C P L R A A V T P A A G V C A R E K P Q G S T V A A P E E E D T D P R R L V Q L L R Q H S S P W Q V Y G F V R A C I L R R L

V P P G L W G S

R H N E R R F L R N T K K F I S L G K H A K L S I Q E L T W K M S V R G C A W L R R S P G V G C V P A A E H R L R E E I L A K F L H W L M S V Y  
V V E L L R S F

F Y V T E T T F Q K N R L F F Y R K S V W S K L Q S I G I R Q H L K R V Q L R E L S E A E V R Q H R E A R P A L L T S R L R F I P K P D G L R P I V N  
M D Y V V

G A R T F R E K R A E R I L T S R V K A L F S V I N Y E R A R R P G L L G A S V U L G L D D I H R A W R T F V L R V R A Q D P P P E L Y F V K V D  
V T G A Y D T I

P O D R I T E V I A S I I K P Q N T Y C V R R Y A V V Q K A A H G H V R K A F K S H V S T I L D I Q P Y M R Q F V A H L Q E T S P L R D A V V I E  
Q S S S L N E

A S S G L F D V F L R F M C H E H A V R I R G K S Y V Q C Q G I P Q G S I I S T I L C S I C Y G D M E N K L F A G I R D G L I I L R L V D D F L L V T  
P H L T H A

K T P F L R T L V R G V P E Y G C V V N L R K T V V N F P V E D E A L G G T A F V Q M P A H G L F P W C G L I I I D T R T L E V Q S D Y S S Y A R T  
S I P A S L T F

N R G F K A G R N M R R K L F G V L R L K H S L F L D L Q V N S L Q T V C T N I Y K I I I L Q A Y R F H A C V L Q L P F H Q Q V W K N P T F F  
I R V I S D T A

S I L C Y S I L K A K N A G M S I L G A K G A A G P L P S E A V Q W L C H Q A F L L K L T R H R V T T V P L I G S L R T A Q T O L S R K L P G T T I T T

A L E A A R N  
P A L P S D F K T I L D

FIG. 6

10/14

10	20	30	40	
GTCTCACTCTGTCACCCAGGCTGGAGTGTAAATGGCACAAAT			40	
CTCGGCTCACTGCAACCTCTGCCTCCTCGGTTCAAGCAGT			80	
TCTCATTCCTAACCTCATGAGTAGCTGGGATTACAGGCG			120	
CCCACCACACGCCCTGGTTAATTTTGATTTTAGTAGA			160	
GATAGGTTTACCATGTTGCCAGGCTGGCTCAAACCTC			200	
210	220	230	240	
CTGACACTAAGTGTATCGCCCGCCTGGCCTCCACAGTG			240	
CTGGGATTACAGGTGCAAGGCCACCGTGCCCGCATACCTT			280	
GATCTTTAAAATGAAGTCTGAAACATTGCTACCCCTGTC			320	
CTGAGGAATAAGACCCCTAGTGATTTAGCTCTGGCAC			360	
CCCCCAGCCTGTTGCTGTTCCCTGCTGACTTAGTTCTA			400	
410	420	430	440	
TCTCAGGCATCTGACACCCCCACAAGCTAAGCAATTATTA			440	
ATATTGTTTCCCGTGTGAGTGTCTGTAGCTTGCCCC			480	
CGCCCTGCTTTCCCTCTGTTCCCGTCTGCTTCTGT			520	
CTCAGGCCCCGCCGCTGGGGTCCCTCTGTCCTTGC			560	
GTGGTTCTCTGCTTGTATGCTGGTAAACCCAGCTT			600	
610	620	630	640	
TACCTGTGCTGGCCTCCATGGCATCTAGCGACGTCGGGG			640	
ACCTCTGCTTATGATGCACAGATGAAGATGTGGAGACTCA			680	
CGAGGAGGGCGGTCACTTGGCCCGTGAGTGTCTGGAGCA			720	
CCACGTGGCCAGCGTCCCTAGCCAGTGAGTGCACAGCAAC			760	
GTCCGCTCGGCTGGGTCAGCCTGGAAAACCCAGGCAN			800	
810	820	830	840	
GTCGGGGCTGGGTCTCCGGGTGTCGAGTTGAAATCG			840	
CGCAAACCTGCGGTGTCGGGCCAGCTCTGACGGTGTGCC			880	
TGGCGGGGGAGNGTCTGCTTCCCTCTGCTTGGAAC			920	
CAGGACAAAGGATGAGGCTCCGAGCCGTTGTCGCCAAC			960	
GGAGCATGACGGGTTGGCTGTGTTCCGGCCGAGAGCACC			1000	
1010	1020	1030	1040	
GTCTCGTGAGGAGATCCTGGCCAAGTCTCTGCACTGGCT			1040	
GATGAGTGTGTACGTCGAGCTGCTAGGTCTTCTT			1080	
TATGTCACGGAGACCACGTTCAAAAGAACAGGCTCTTT			1120	
TCTACCGGAAGAGTGTCTGGAGCAAGTTGCAAAGCATTGG			1160	
AATCAGACAGCACTGAAAGAGGGTGCAGCTGCGGGAGCTG			1200	

FIG. 7A

11/14

1210	1220	1230	1240	
TCGGAAGCAGAGGTCAAGGCAGCATCGGAAGCCAGGCCCG		1240		
CCCTGCTGACGTCCAGACTCCGCTTCATCCCCAAGCCTGA		1280		
CGGGCTGCGCCGATTGTGAACATGGACTACGTGCTGGGA		1320		
GCCAGAACGTTCCGAGAGAAAAGAGGGCCGAGCGTCTCA		1360		
CCTCGAGGGTGAAGGCACTGTTCAAGCGTGCTCAACTACGA		1400		
1410	1420	1430	1440	
GCGGGCGCGGCCGCCGGCTCCTGGGCGCTCTGTGCTG		1440		
GGCCTGGACGATATCCACAGGGCCTGGCGCACCTTCGTGC		1480		
TGCGTGTGCGGGCCCAGGACCCGCGCCTGAGCTGTACTT		1520		
TGTCAAGGTGGATGTGACGGCGCGTACGACACCATCCCC		1560		
CAGGACAGGCTACGGAGGTATCGCCAGCATCATCAAAC		1600		
1610	1620	1630	1640	
CCCAGAACACGTACTGCGTGCCTCGGTATGCCGTGGTCCA		1640		
GAAGGGCGCCCATGGGACGTCCCGCAAGGCCTTCAAGAGC		1680		
CACGTCCTACGTCAGTGCAGGGGATCCCGCAGGGCTCC		1720		
ATCCTCTCACGCTGCTCTGCAGCCTGTGCTACGGCGACA		1760		
TGGAGAACAGCTGTTGCAGGGATTCCGGGGACGGGCT		1800		
1810	1820	1830	1840	
GCTCCTGCGTTGGTGGATGATTCTTGTGGTGACACCT		1840		
CACCTCACCCACCGGAAAACCTTCCTCAGGACCCCTGGTCC		1880		
GAGGTGTCCTGAGTATGGCTGCGTGGTGAACCTGGGAA		1920		
GACAGTGGTGAACCTCCCTGAGAACGAGGCCCTGGGT		1960		
GGCACGGCTTGTTCAGATGCCGGCCACGGCCTATTCC		2000		
2010	2020	2030	2040	
CCTGGTGCAGGCCCTGCTGCTGGATAACCGGACCCCTGGAGGT		2040		
GCAGAGCGACTACTCCAGCTATGCCGGACCTCCATCAGA		2080		
GCCAGTCTCACCTCAACCGCGGCTTCAGGCTGGGAGGA		2120		
ACATGCGTGCACACTCTTGGGGTCTTGCGGCTGAAGTG		2160		
TCACAGCCTTTCTGGATTGCAAGGTGAACAGCCTCCAG		2200		
2210	2220	2230	2240	
ACGGTGTGACCAACATCTACAAGATCCTCCTGCTGCAGG		2240		
CGTACAGGTTTACCGCATGTTGCTGCAGCTCCATTCA		2280		
TCAGCAAGTTGGAGAACCCCACATTTCCTGCGCGTC		2320		
ATCTCTGACACGGCCTCCCTGCTACTCCATCCTGAAAG		2360		
CCAAGAACGCAAGAACATTTCTGCGTGAACAGCCTCCAG		2400		
2410	2420	2430	2440	
TGCGGTGCTTGGGTGGGACAGCCAGAGATGGAGGCCACCC		2440		
CGCAGACCGTGGGTGGGAGCTTCCGGTGTCTCCTG		2480		
GGAGGGGAGCTGGGCTGGGCTGTGACTCCTCAGCCTCTG		2520		
TTTCCCCCAGGGATGTCGCTGGGGCCAAGGGCCGCC		2560		
GGCCCTCTGCCCTCCGAGGCCGTGCAGTGGCTGTGCCACC		2600		

FIG. 7B

12/14

2610	2620	2630	2640
AAGCATTCCCTGCTCAAGCTGACTCGACACCGTGTACCTA			2640
CGTGCCACTCCCTGGGGTCACTCAGGACAGCCCAGCGAG			2680
CTGAGTCGGAAGCTCCGGGACGACGCTGACTGCCCTGG			2720
AGGCCGCAGCCAACCCGGCACTGCCCTCAGACTCAAGAC			2760
CATCCTGGACTGATGCCACCCGCCACAGCCAGGCCAG			2800

2810	2820	2830	2840
AGCAGACACCAGCAGCCCTGTCACGCCGGCTCACGTCC			2840
CAGGGAGGGAGGGGCGGCCACACCCAGGCCGACCGCT			2880
GGGAGTCTGAGGCCCTGAGTGAGTGTTTGGCCGAGGCCTGC			2920
ATGTCGGCTGAAGGCTGAGTGTCGGCTGAGGCCTGAGC			2960
GAGTGTCCAGCCAAGGGCTGAGTGTCAGCACACCTGCCG			3000

3010	3020	3030	3040
TCTTCACCTCCCCACAGGCTGGCGCTCGGCTCCACCCCAG			3040
GGCCAGCTTTCCTCACCAAGGAGCCGGCTTCCACTCCCC			3080
ACATAGGAATAGTCCATCCCAGATTGCCATTGTTCCACC			3120
CCTGCCCTGCCCTCCCTTGCCCTCCACCCCCACCATCCA			3160
GGTGGAGACCTGAGAAGGACCTGGGAGCTCTGGGAATT			3200

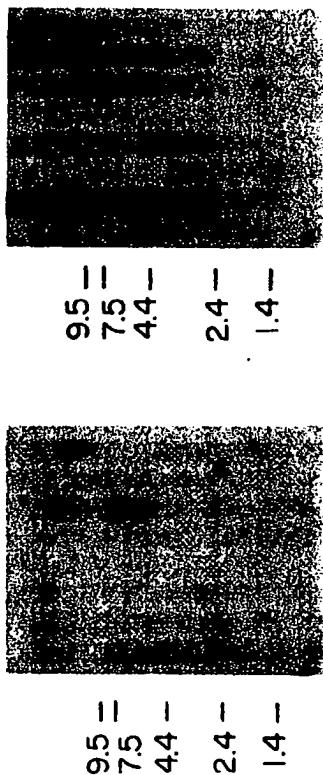
3210	3220	3230	3240
TGGAGTGACCAAAGGTGTGCCCTGTACACAGGCAGGGACC			3240
CTGCACCTGGATGGGGTCCCTGTGGGTCAAATTGGGGGG			3280
AGGTGCTGTGGGAGTAAATACTGAATATATGAGTTTTC			3320
AGTTTGAAAAAAAAAGGAATTTC 3346			

FIG. 7C

Normal Human Tissues

FIG. 8A

heart  
brain  
placenta  
lung  
liver  
muscle  
kidney  
pancreas



Cancer Cell Lines

FIG. 8C

HL-60  
K-562  
HeLa  
Raji  
MOLT-4  
SW480  
A549  
G361



FIG. 8B

spleen  
thyroid  
prostate  
testis  
ovary  
colon  
small intestine  
leukocyte

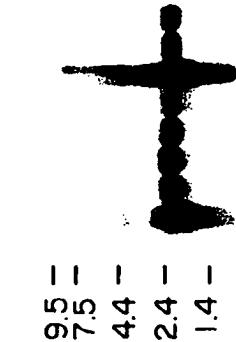
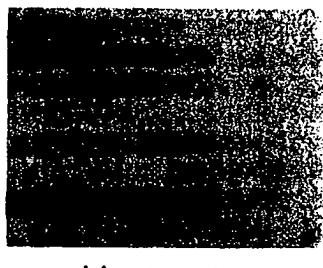


FIG. 8E

FIG. 8F

14/14

FIG. 9A

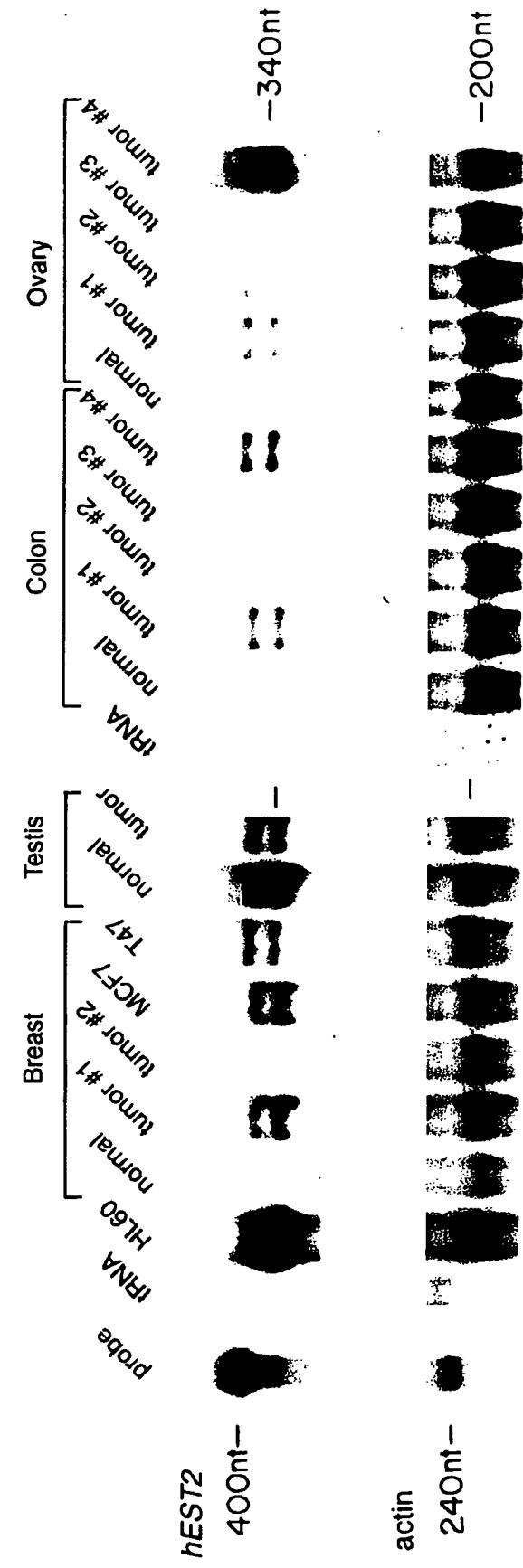


FIG. 9C

FIG. 9D

FIG. 9B

